

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2016.057a-dB			(to be completed by ICTV officers)			
Short title: To create one (1) new genus, <i>Kpj</i> family <i>Podoviridae</i> . (e.g. 6 new species in the genus <i>Zetavirus</i>) Modules attached (modules 1 and 10 are required)		25virus, i 1 ⊠ 6 □	2 ⊠ 7 □	3 ⊠ 8 □	ew species in	5 □ 10 ⊠	
Author(s):							
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Andrew M. Kropinski Phage.C	Canada@gmail.	com					
List the ICTV study group(s)	that have see	n this pro	posal:				
A list of study groups and contacts is provided at http://www.ictvonline.org/subcommittees.asp . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses) ICTV Bacterial and Archaeal Vi Subcommittee					Viruses		
ICTV Study Group comments (if any) and response of the proposer:							
Date first submitted to ICTV: Date of this revision (if different	eate first submitted to ICTV: June 2016 eate of this revision (if different to above):						
ICTV-EC comments and response of the proposer:							

MODULE 2: NEW SPECIES

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	Code $2016.057aB$ (assigned by IC			ICTV office	ers)			
To crea	ite 2 ne	ew species with	in:					
		,			Fill in all that apply. • If the higher taxon has yet to be			
	Genus:	<i>Kpp25virus</i> (n	ew)					
Subfamily:					ated (in a later module, below) write ew)" after its proposed name.			
Fa	amily:	mily: Podoviridae		•	If no genus is specified, enter			
(Order:	Caudovirales			"unassigned" in the genus box.			
Name o	of new	species:	Representative isol 1 per species please)	ate: (only	GenBank sequence accession number(s)			
Pseudomonas virus KPP25 Pseudo		Pseudomonas phage	KPP25	AB910393				
Pseudomonas virus R18		Pseudomonas phage	PhiR18	LC102729				

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, explain how the new species meet these criteria.
 - o If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 3: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	201	6.057bB	(assigned by ICTV officers)			
To create	a new	genus within:		Fill in all that apply.		
Subfa	mily:			If the higher taxon has yet to be created		
Far	mily:	Podoviridae		(in a later module, below) write "(new)" after its proposed name.		
0	rder:	Caudovirales		 If no family is specified, enter "unassigned" in the family box 		

naming a new genus

Code	2016.057cB	(assigned by ICTV officers)
To name the	he new genus: Kpp25virus	

Assigning the type species and other species to a new genus

Code	2016.057dB	(assigned by ICTV officers)					
To designate the following as the type species of the new genus							
Pseudomonas virus KPP25 Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered							
The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). Please enter here the TOTAL number of species (including the type species) that the genus will contain:							

Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

Pseudomonas aeruginosa phages KPP25 [5] and ΦR18 [4] were isolated from an agricultural wastewater drain in Kochi City (Japan) and, from sewage treatment plants at Hokkaido (Japan). In the case of KPP25 the isometric head diameter and tail length are 63 and 30 nm, respectively (Fig. 1). The authors [5] state that "phage KPP25 differed from the known *Pseudomonas* podophages belonging to the genera of phiKMV-like, T7-like, N4-like, and Luz24-like viruses, as well as the unclassified *Pseudomonas* podophages." The absence of integrases or recombinases suggests that these are lytic phages.

NCBI BLASTN, CoreGenes (Table 1) [2], progressiveMauve [1] (Fig. 2), and phylogenetic analyses (Fig. 3) [3] all indicate that the proposed genus, *Kpp25virus*, is cohesive and distinct from other genera. On average, the genomes of this genus are 63.8 kb in length (60.3 mol% G+C), and encode 87 proteins and 0 tRNAs.

Origin of the new genus name:

Based upon the name of the first sequenced member of this genus.

Reasons to justify the choice of type species:

The first sequenced member of this genus.

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm

MODULE 10: **APPENDIX**: supporting material

additional material in support of this proposal

References:

- 1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.
- 2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes 3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.
- 3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.
- 4. Agren J, Sundström A, Håfström T, Segerman B. Gegenees: fragmented alignment of multiple genomes for determining phylogenomic distances and genetic signatures unique for specified target groups. PLoS One. 2012;7(6):e39107.
- 5. Miyata R, Yamaguchi K, Uchiyama J, Shigehisa R, Takemura-Uchiyama I, Kato S, Ujihara T, Sakaguchi Y, Daibata M, Matsuzaki S. Characterization of a novel *Pseudomonas aeruginosa* bacteriophage, KPP25, of the family *Podoviridae*. Virus Res. 2014;189:43-6.
- 6. Furusawa T, Iwano H, Higuchi H, Usui M, Maruyama F, Nakagawa I, Yokota H, Tamura Y. Complete Genome Sequences of Broad-Host-Range *Pseudomonas aeruginosa* Bacteriophages ΦR18 and ΦS12-1. Genome Announc. 2016;4(3). pii: e00041-16.

Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

Table 1. Properties of the phages belonging to the genus *Kpp25virus*.

Pseudomonas	RefSeq No.	GenBank	Genome	Genome	No.	% DNA	%
phage	_	Accession	length	(mol%	CDS	Sequence	Homologous
		No.	(kb)	G+C)		identity *	proteins **
KPP25	NC_024123	AB910393	64.11	60.3	88	100	100
PhiR18		LC102729	63.56	60.3	86	94	94.3

^{*} Determined using BLASTN; ** Determined using CoreGenes [2].

Fig. 1. Electron micrograph of negatively stained Pseudomonas phage KPP25 [5] - Permission provided by: Dr. Jumpei Uchiyama, Department of Veterinary Microbiology, School of Veterinary Medicine, Azabu University, Fuchinobe 1-7-71, Chuo-ku Sagamihara-shi, Kanagawa 252-0206, Japan.

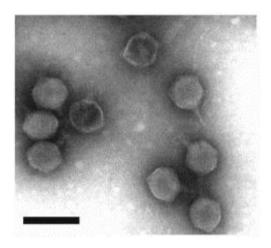


Fig. 2. progressiveMauve alignment [1] of the genomes of members of the *Kpp25virus* genus – from top to bottom: Pseudomonas phages KPP25 and Phi18. Colored blocks indicate the regions of 1 to 1 best alignment with rearrangement breakpoints in a different random color. The degree of sequence similarity between regions is given by a similarity plot within the colored blocks with the height of the plot proportional to the average nucleotide identity (Aaron Darling, personal communication). N.B. The two genomes are not collinear.

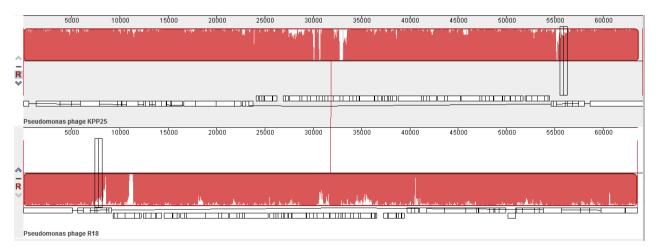


Fig. 3. Phylogenetic analysis of (A) major capsid protein and (B) terminase large subunit (TerL) proteins of KPP25-like viruses and homologous proteins from a variety of other phages constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."

A. Major capsid protein

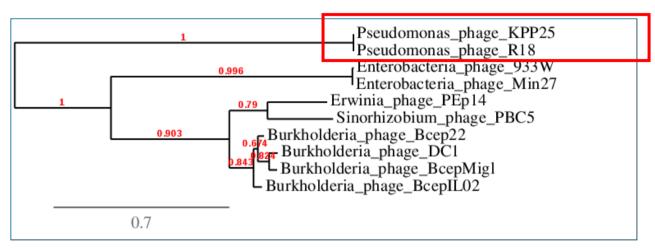


Figure 1: Phylogenetic tree (the branch length is proportional to the number of substitutions per site).

B. TerL proteins

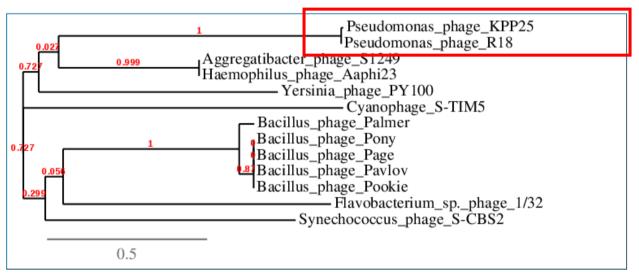


Figure 1: Phylogenetic tree (the branch length is proportional to the number of substitutions per site).