MODULE 1: **TITLE, AUTHORS, etc**

Code assigned:	*2016.033a-dB*	(to be completed by ICTV officers)

Short title: To create one (1) new genus, *P12024virus*, including two (2) new species in the family *Siphoviridae*.
(e.g. 6 new species in the genus *Zetavirus*)

| **Modules attached** (modules 1 and 10 are required) | 1 ☒ | 2 ☒ | 3 ☒ | 4 ☐ | 5 ☐ |
| | 6 ☐ | 7 ☐ | 8 ☐ | 9 ☐ | 10 ☒ |

Author(s):

Jang-Cheon Cho— Inha University (Republic of Korea)
Jens H. Kuhn—NIH/NIAID/IRF-Frederick, Maryland (USA)
Andrew M. Kropinski—University of Guelph (Canada)
Evelien M. Adriaenssens—University of Pretoria (South Africa)

Corresponding author with e-mail address:

Andrew M. Kropinski Phage.Canada@gmail.com

List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided at http://www.ictvonline.org/subcommittees.asp . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses)	ICTV Bacterial and Archaeal Viruses Subcommittee

ICTV Study Group comments (if any) and response of the proposer:

Date first submitted to ICTV:	June 2016
Date of this revision (if different to above):	

ICTV-EC comments and response of the proposer:

MODULE 2: **NEW SPECIES**

creating and naming one or more new species.
If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	*2016.033aB*		(assigned by ICTV officers)

To create 2 new species within:

Genus:	*P12024virus* **(new)**	Fill in all that apply.
Subfamily:		• If the higher taxon has yet to be created (in a later module, below) write "**(new)**" after its proposed name.
Family:	*Siphoviridae*	• If no genus is specified, enter "**unassigned**" in the genus box.
Order:	*Caudovirales*	

Name of new species:	Representative isolate: (only 1 per species please)	GenBank sequence accession number(s)
Nonlabens virus P12024S *Nonlabens virus P12024L*	Nonlabens phage P12024S Nonlabens phage P12024S	JQ823122.1 JQ823123.1

Reasons to justify the creation and assignment of the new species:
- Explain how the proposed species differ(s) from all existing species.
  - If species demarcation criteria (see module 3) have previously been defined for the genus, **explain how the new species meet these criteria**.
  - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 3: **NEW GENUS**

Ideally, a genus should be placed within a higher taxon.

Code	*2016.033bB*	(assigned by ICTV officers)

To create a new genus within:

Fill in all that apply.
- If the higher taxon has yet to be created (in a later module, below) write "**(new)**" after its proposed name.
- If no family is specified, enter "**unassigned**" in the family box

Subfamily:	
Family:	*Siphoviridae*
Order:	*Caudovirales*

naming a new genus

Code	*2016.033cB*	(assigned by ICTV officers)

To name the new genus: *P12024virus*

Assigning the type species and other species to a new genus

Code	*2016.033dB*	(assigned by ICTV officers)

To designate the following as the type species of the new genus

Nonlabens virus P12024S

Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered

The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). **Please enter here the TOTAL number of species (including the type species) that the genus will contain:**

2

Reasons to justify the creation of a new genus:
Additional material in support of this proposal may be presented in the Appendix, Module 9

When these two phages were first characterized, the host was defined as a marine bacterium of the genus *Persicivirga* (family *Flavobacteriaceae*, *Bacteroidetes* phylum). This genus contains members able to degrade complex polysaccharides, such as xylan and ulvan [4]. Subsequent research indicated that this genus is synonymous with *Nonlabens* [5]. There is no notable difference in size between the two phages. The head diameters are approximately 47 nm and tail lengths are ~90 nm. The phage particles display a prominent collar.

BLASTN, CoreGenes (Table 1) [2], progressiveMauve alignment (Fig. 2) [1], and phylogenetic analyses (Fig. 3) [3] all indicate that the proposed genus, *P12024virus*, is cohesive and distinct from other genera. On average, the genomes of this genus are 35.7 kb in length (35.7 mol% G+C), and encode 58-59 proteins and 0 tRNAs.

Origin of the new genus name:

Based upon the name of the first sequenced member of this genus.

Reasons to justify the choice of type species:

The first sequenced member of this genus.

Species demarcation criteria in the new genus:

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 10: **APPENDIX**: supporting material

additional material in support of this proposal

References:

1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.

2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.

3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.

4. Kang I, Jang H, Cho JC. Complete genome sequences of two *Persicivirga* bacteriophages, P12024S and P12024L. J Virol. 2012;86(16):8907-8.

5. Yi H, Chun J. Unification of the genera *Nonlabens, Persicivirga, Sandarakinotalea* and *Stenothermobacter* into a single emended genus, *Nonlabens*, and description of *Nonlabens agnitus* sp. nov. Syst Appl Microbiol. 2012;35(3):150-5.

Annex:

Fig. 1. Electron micrograph of negatively stained Nonlabens phages P12024S (left) and P12024L (right).
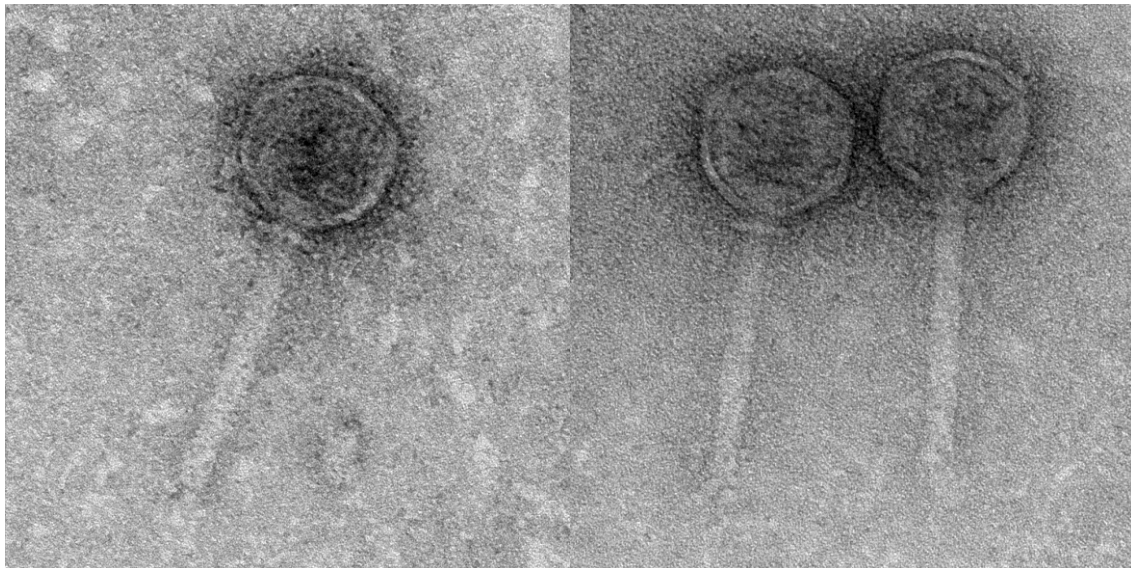
Table 1. Properties of the two phages belonging to the genus *P12024virus*.

Nonlabens phage	RefSeq No.	GenBank Accession No.	Genome length (kb)	Genome (mol% G+C)	No. CDS	DNA (% sequence identity)*	% Homologous proteins **
P12024S	NC_018271.1	JQ823122.1	35.70	35.5	59	100	100
P12024L	NC_018272.1	JQ823123.1	35.65	35.9	58	88	89.8

* Determined using BLASTN; ** Determined using CoreGenes [2]

Fig. 2. progressiveMauve alignment [1] of the annotated genomes of members of the *P12024virus* genus – from top to bottom: Nonlabens phages P12024S and P12024L. Colored blocks indicate the regions of 1 to 1 best alignment with rearrangement breakpoints in a different random color. The degree of sequence similarity between regions is given by a similarity plot within the colored blocks with the height of the plot proportional to the average nucleotide identity (Aaron Darling, personal communication).
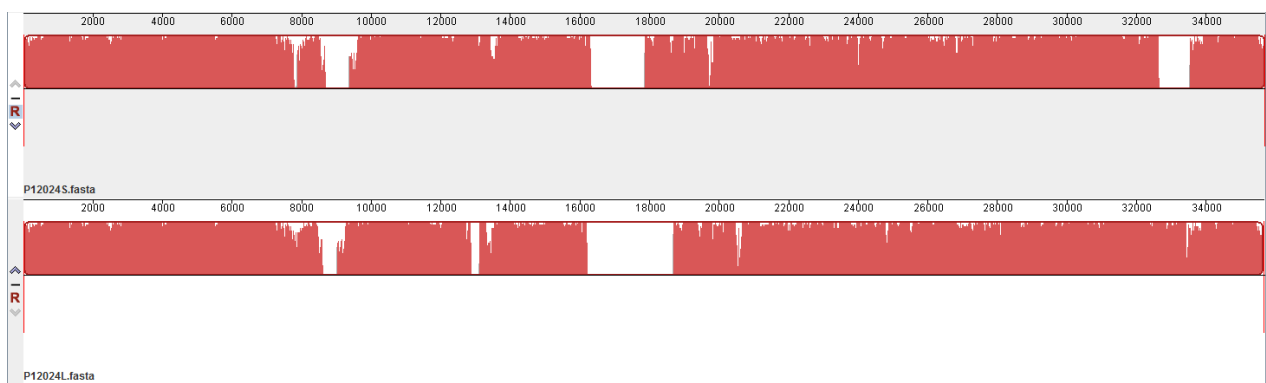
Fig. 3. Phylogenetic analysis of large subunit terminase proteins of Nonlabens phage P12014-like viruses and homologous proteins from a variety of other phages constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."
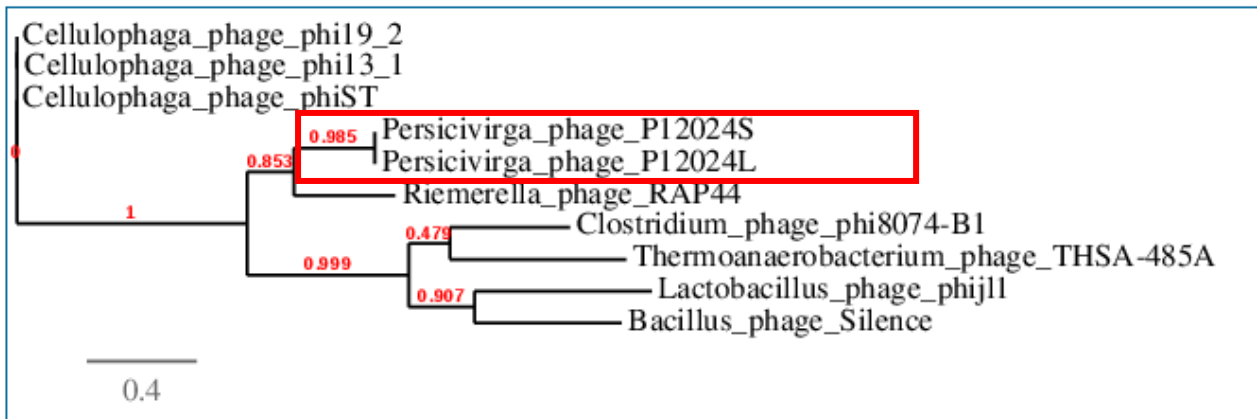

Figure 1: *Phylogenetic tree (the branch length is proportional to the number of substitutions per site).*