

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

# MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2013.034a-dB			(to be completed by ICTV officers)			
Short title: To create a new genus, the <i>Sfi1unalikevirus</i> , within the family <i>Siphoviridae</i> (e.g. 6 new species in the genus <i>Zetavirus</i> )							
Modules attached (modules 1 and 9 are required)		1 ⊠ 6 □	2 × 7 □	3 ⊠ 8 □	4 □ 9 ⊠	5 🗌	
Author(s) with e-mail address	ss(es) of the pr	oposer:					
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List the ICTV study group(s) that have seen this proposal:							
A list of study groups and contact							
http://www.ictvonline.org/subcom in doubt, contact the appropriate							
chair (fungal, invertebrate, plant, prokaryote or							
vertebrate viruses)							
ICTV-EC or Study Group comments and response of the proposer:							
Date first submitted to ICTV:			Iuma	2013			
Date of this revision (if differe	nt to above).			2013 2014			

# **MODULE 2: NEW SPECIES**

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for one isolate of each new species proposed.

Code 2	01.	3.034aB	(assigned by IC	CTV offic	cers)		
To create 5 new species within:					Fill in all that apply.		
Genus: Sfi1unalikevirus (new)			<ul> <li>If the higher taxon has yet to be created (in a later module, below) write "(new)" after its proposed name.</li> </ul>				
Subfamily:							
Famil	Family: Siphoviridae		If no genus is specified, enter				
Orde	er:	Caudovirales	"unassigned" in the genus box.				
					GenBank sequence accession number(s) of reference isolate:		
Streptocoo	сси	s phage Sfi11			AF158600		
Streptocoo	сси	s phage 01205		U88974			
Streptocoo	сси	s phage 2972			AY699705		
Streptococcus phage Alq132				FJ226752			
Streptocoo	CCU.	s phage 858			EF529515		

# Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
  - o If species demarcation criteria (see module 3) have previously been defined for the genus, **explain how the new species meet these criteria**.
  - o If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

BLASTN analyses reveal that these five *Streptococcus* phages are related and distinct from any other phage. We have chosen 95% DNA sequence identity as the criterion for demarcation of species.

# **MODULE 3: NEW GENUS**

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	201	3.034bB	(assigned by IC	CTV officers)
To create a	a new	genus within:		Fill in all that apply
0.10	•1			<ul><li>Fill in all that apply.</li><li>If the higher taxon has yet to be created</li></ul>
Subfar	nıly:			(in a later module, below) write "(new)"
Far	nily:	Siphoviridae		after its proposed name.
O	rder:	Caudovirales		If no family is specified, enter
				"unassigned" in the family box

naming a new genus

Code	2013.034cB	(assigned by ICTV officers)				
To name the new genus: Sfi1unalikevirus						

Assigning the type species and other species to a new genus

Code	2013.034dB	(assigned by ICTV officers)					
To design	To designate the following as the type species of the new genus						
Streptococcus phage Sfi11  Every genus must have a type species. This sh be a well characterized species although not necessarily the first to be discovered							
The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). Please enter here the TOTAL number of species (including the type species) that the genus will contain:  5							

#### Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

Phages belonging to this genus have small, isometric heads, and long non-contractile tails. Phages share a comparable genome size and GC content, and a common packaging mechanism with *pac* sites [1–6] (Table 1).

We propose 40% shared proteins with the type phage for new phages to be included in the new genus, as calculated with CoreGenes [7–9].

# Origin of the new genus name:

Streptococcus phage sf11

# Reasons to justify the choice of type species:

One of the first isolates of this group, a well-characterized phage.

# Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. Each of the proposed species differs from the others with more than 5% at the DNA level as confirmed with the EMBOSS Stretcher algorithm, with the two most related phages ALQ13.2 and 2972 sharing 71.5% DNA identity.

# MODULE 9: APPENDIX: supporting material

additional material in support of this proposal

#### **References:**

- Stanley E, Fitzgerald GF, Le Marrec C, Fayard B, van Sinderen D (1997) Sequence analysis and characterization of phi O1205, a temperate bacteriophage infecting *Streptococcus thermophilus* CNRZ1205. Microbiology 143: 3417–3429.
- 2. Guglielmotti DM, Deveau H, Binetti AG, Reinheimer JA, Moineau S, et al. (2009) Genome analysis of two virulent *Streptococcus thermophilus* phages isolated in Argentina. Int J Food Microbiol 136: 101–109. doi:10.1016/i.iifoodmicro.2009.09.005.
- 3. Deveau H, Barrangou R, Garneau JE, Labonté J, Fremaux C, et al. (2008) Phage response to CRISPR-encoded resistance in *Streptococcus thermophilus*. J Bacteriol 190: 1390–1400. doi:10.1128/JB.01412-07.
- 4. Proux C, van Sinderen D, Suarez J, Garcia P, Ladero V, et al. (2002) The dilemma of phage taxonomy illustrated by comparative genomics of Sfi21-like *Siphoviridae* in lactic acid bacteria. J Bacteriol 184: 6026–6036. doi:10.1128/JB.184.21.6026-6036.2002.
- 5. Lucchini S, Desiere F, Brüssow H (1999) Comparative genomics of *Streptococcus thermophilus* phage species supports a modular evolution theory. J Virol 73: 8647–8656.
- Lévesque C, Duplessis M, Labonté J, Labrie S, Fremaux C, et al. (2005)
   Genomic organization and molecular analysis of virulent bacteriophage 2972
   infecting an exopolysaccharide-producing Streptococcus thermophilus strain.
   Appl Environ Microbiol 71: 4057–4068. doi:10.1128/AEM.71.7.4057 4068.2005.
- 7. Mahadevan P, King JF, Seto D (2009) CGUG: in silico proteome and genome parsing tool for the determination of "core" and unique genes in the analysis of genomes up to ca. 1.9 Mb. BMC Res Notes 2: 168. doi:10.1186/1756-0500-2-168.
- 8. Mahadevan P, King JF, Seto (2009) Data mining pathogen genomes using GeneOrder and CoreGenes and CGUG: gene order, synteny and in silico proteomes. Int J Comput Biol Drug Des 2: 100–114.
- 9. Zafar N, Mazumder R, Seto D (2002) CoreGenes: A computational tool for identifying and cataloging "core" genes in a set of small genomes. BMC Bioinformatics 3: 12. doi:10.1186/1471-2105-3-12.
- 10. Darling AE, Mau B, Perna NT (2010) progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5: e11147.

additional material in support of this proposal

### **References:**

doi:10.1371/journal.pone.0011147.

#### Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

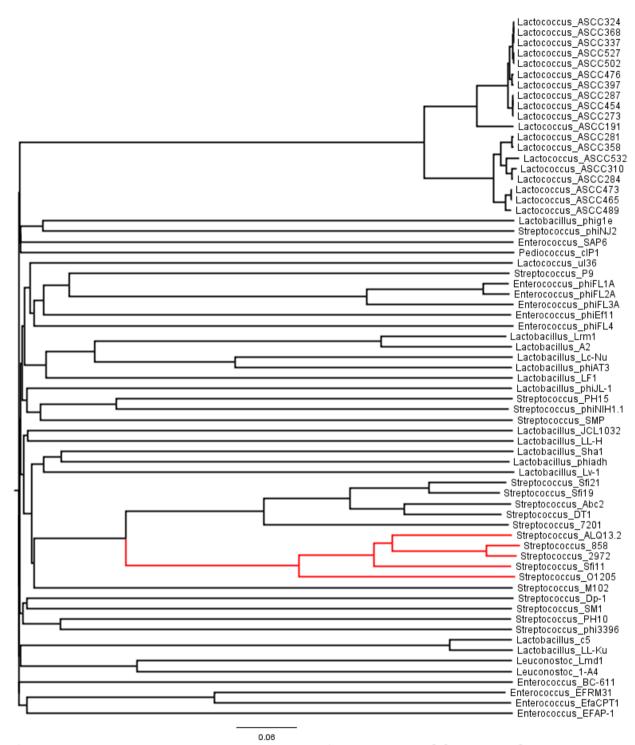
Table 1. Phage genomes

Phage	Accession No.	Size (bp)	Mol%G+C	Packaging – termini	% DNA sequence relatedness (a)	% Protein relatedness (b)
Sfi11	AF158600	39807	39	pac site	100	100
O1205	U88974	43075	38	pac site	ND	92.2
2972	AY699705	34704	40	pac site	67.6	70.6
Alq13.2	FJ226752	35525	39	pac site	66.0	66.7
858	EF529515	35543	40	pac site	65.8	74.5

- (a) EMBOSS Stretcher values relative to the type phage Sfi11 could not be calculated for O1205 due to different origins of the sequences in GenBank
- (b) CoreGenes 3.5 (relative to Sfi11)



**Figure 1.** progressiveMauve alignment of the genomes of the these *Streptococcus* phages indicates that they are very similar [10]. The sequence of phage O1205 was deposited in GenBank with a different starting point from the others. Colored blocks indicate the regions of 1 to 1 best alignment with rearrangement breakpoints in a different random color. The degree of sequence similarity between regions is given by a similarity plot within the colored blocks with the height of the plot proportional to the average nucleotide identity.



**Figure 2:** Whole genome phylogenetic tree of selected low GC content *Siphoviridae* phages in the NCBI database in November 2012. Genome sequences were aligned and the tree (NJ) was created with ClustalW 2.0 and visualized with FigTree. The newly proposed genus is colored in red.