

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2015.001a-kF		(to be cor officers)	mpleted by I	CTV
Short title: A new family and two new genera for classification of virophages					
two new species (e.g. 6 new species in the genus a Modules attached (modules 1 and 10 are required)	Zetavirus) 1 🔀 6 🗖	2 🔀 7 🗌	3 × 8 □	4 9	5 🖂 10 🖂

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List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided	
at http://www.ictvonline.org/subcommittees.asp	
. If in doubt, contact the appropriate	
subcommittee chair (fungal, invertebrate, plant,	
prokaryote or vertebrate viruses)	

ICTV Study Group comments (if any) and response of the proposer:

Date first submitted to ICTV: Date of this revision (if different to above): June 11, 2015

ICTV-EC comments and response of the proposer:

Fungal and Protist Viruses Subcommittee Chair: Proposal approved for submission.

MODULE 2a: **NEW SPECIES**

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	201	5.001aF	(assigned by IC	TV officers)	
To crea	te 2 ne	ew species within:			
				Fill in all that a	oply.
G	lenus:	Sputnikvirus (new))	 If the higher t 	axon has yet to be
Subfa	mily:			"(new)" after	its proposed name
Fa	mily:	Lavidaviridae (new	v)	 If no genus is 	specified, enter
(Order:			"unassigned	" in the genus box.
Name o	of new a	species:	Representative is per species please)	olate: (only 1	GenBank sequence accession number(s)
Mimivir	·us-dep	endent virus	Sputnik virus 1		EU606015
Sputnik Mimivirus-dependent virus Zamilon		Zamilon virus		HG531932	

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, **explain how the new species meet these criteria**.
 - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
 - Further material in support of this proposal may be presented in the Appendix, Module 9

Both virus species represent satellite-like viruses of protists, also known as virophages, which depend for multiplication on members of the genus *Mimivirus* within the family *Mimiviridae*. Sputnik and Zamilon are able to replicate in *Acanthamoeba polyphaga* cells that are co-infected with Acanthamoeba polyphaga mimivirus (APMV) or Mont1 mimivirus, respectively [1, 2]. The latter virus is more closely related to "*Megavirus chilensis*" [3] than to APMV [4]. Both Sputnik and Zamilon have similarly-sized (17,276-18,342 base pairs), circular double-stranded DNA genomes that code for 20-21 proteins (Module 10; Figure 1) [1, 2, 5, 6]. The virions are icosahedral with a diameter of 60-75 nm, and are composed of at least two different proteins with jelly-roll folds, the minor and the major capsid protein (Module 10; Figure 2a) [7, 8]. The capsid proteins of viruses in the proposed genus are homologous and the major capsid protein can be used as a phylogenetic marker to demonstrate membership in the proposed genus (Module 10; Figure 3).

In addition to the two capsid protein genes, both species encode a FtsK-HerA family DNApackaging ATPase, a cysteine protease, a primase-superfamily 3 helicase, a lambda-type integrase, a transposase, a Zinc-ribbon domain protein, a collagen-like protein, and six proteins of unknown function [1, 2].

MODULE 2b: NEW SPECIES

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	201	5.001bF (assigned by IC		TV officers)	
To create 1 new species within:			:		
				Fill in all th	nat apply.
Genus: Mavirus (new)			If the higher taxon has yet to be		
Subfa	mily:			created (in a later module, below) write "(new)" after its proposed name	
Fa	Family: <i>Lavidaviridae</i> (new)		 If no genus is specified, enter 		
C	Order:	r:		"unassigned" in the genus box.	
Name of new species: Re per		Representative isola per species please)	te: (only 1	GenBank sequence accession number(s)	
Cafeteriavirus-dependent Mave		Maverick-related viru	18	HQ712116	
mavirus (Mavi		(Mavirus), strain Sper	zl		

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, explain how the new species meet these criteria.
 - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

Mavirus replicates in the marine heterotrophic nanoflagellate *Cafeteria roenbergensis* in the presence of Cafeteria roenbergensis virus (CroV) [9, 10]. The circular, double-stranded DNA genome consists of 19,063 bp and encodes 20 proteins. The virions are icosahedral with a diameter of ~75 nm, and are composed of at least two different proteins with jelly-roll folds, the minor and the major capsid protein (Module 10; Figure 2b). The capsid proteins of viruses in the proposed genus are homologous and the major capsid protein can be used as a phylogenetic marker to demonstrate membership in the proposed genus (Module 10; Figure 3). In addition to the two capsid protein genes, Mavirus encodes a FtsK-HerA family DNApackaging ATPase, a cysteine protease, a superfamily 3 helicase, a retroviral-type integrase, a lipase, and a FNIP repeat-containing protein [9]. Mavirus shares many features with the large, virus-like transposons of the Maverick/Polinton superfamily which are widespread in eukaryotes [11, 12]. Both types of elements encode 7 homologous proteins involved in virion morphogenesis (minor and major capsid proteins, FtsK-HerA-type genome packaging ATPase and cysteine protease homologous to adenoviral maturation proteases), genome replication (protein-primed family B DNA polymerase and superfamily 3 helicase) and integration (retrovirus-like integrase which belongs to a broad superfamily of DDE transposases) [9, 13]. Furthermore, the Mavirus genome contains long inverted repeats that resemble those found at the termini of Maverick/Polinton transposons.

MODULE 3a: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	2015.001cF	(assigned by ICTV officers)
To create	a new genus within:	Fill in all that apply.
Subfa	mily:	 If the higher taxon has yet to be created (in a later module, helper) write "(new)"
Fai	mily: Lavidaviridae (new)	after its proposed name.
0	rder:	 If no family is specified, enter "unassigned" in the family box

naming a new genus

Code	2015.001dF	(assigned by ICTV officers)
To name the new genus: Sputnikvirus		

Assigning the type species and other species to a new genu

Assigning	the type species and other specie	es to a new genus	
Code	2015.001eF	(assigned by ICTV officers)	
To designa	te the following as the type sp	becies of the new genus	
Mimivirus-	dependent virus Sputnik	Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered	
The new ger are being m (including	nus will also contain any other new oved from elsewhere (Module 7b). the type species) that the genu	I species created and assigned to it (Module 2) and any that Please enter here the TOTAL number of species us will contain:	

2

Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

Viruses in this genus depend on members of the genus *Mimivirus* within the family *Mimiviridae* for multiplication and are able to replicate in *Acanthamoeba polyphaga* cells that are co-infected with their respective associated giant virus. All viruses in this genus contain similarly-sized (17,276-18,342 base pairs), circular double-stranded DNA genomes that code for 20-21 proteins. (Module 10; Figure 1) [1, 2, 5, 6]. The virions are icosahedral with a diameter of 60-75 nm, and are composed of at least two different proteins with jelly-roll folds, the minor and the major capsid protein (Module 10; Figure 2a) [7, 8].

In addition to the two capsid protein genes, both species encode a FtsK-HerA family DNApackaging ATPase, a cysteine protease, a primase-superfamily 3 helicase, a lambda-type integrase, a transposase, a Zinc-ribbon domain protein, a collagen-like protein, and six proteins of unknown function [1, 2]. Based on the large number of homologous genes, it can be assumed that Sputnik and Zamilon are derived from a common ancestor and are best assigned within the same genus.

Origin of the new genus name:

Sputnik was the first virophage to be isolated and represents the type species of this genus.

Reasons to justify the choice of type species:

This representative has been well studied and its genome sequence as well as virion structure are available [1, 6, 8, 14, 15].

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

Membership in the genus is based on the dependence on a giant DNA virus of the genus *Mimivirus* within the family *Mimiviridae*, and on the presence of a set of about 15 homologous genes. These include a major and a minor capsid protein gene, a FtsK-HerA family DNA-packaging ATPase, a cysteine protease, a primase-superfamily 3 helicase, and a lambda-type integrase [2]. The capsid proteins of viruses in the proposed genus are homologous and the major capsid protein can be used as a phylogenetic marker to demonstrate membership in the proposed genus (Module 10; Figure 3).

MODULE 3b: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code 20	15.001fF	(assigned by ICTV officers)
To create a nev	w genus within:	
		Fill in all that apply.
Subfamily:		 If the higher taxon has yet to be created (in a later module, helper) write "(new)"
Family:	Lavidaviridae (new)	after its proposed name
Order:		 If no family is specified, enter
		"unassigned" in the family box

naming a new genus

Code	2015.001gF	(assigned by ICTV officers)
To name the new genus: <i>Mavirus</i>		

Assigning the type species and other species to a new genus

1 1001511115	the type species and other speci	ies to a new Senas	
Code	2015.001hF	(assigned by ICTV officers)	
To designa	ate the following as the type s	pecies of the new genus	
Cafeteriavirus-dependent mavirusEvery genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered			
The new ge are being m (including	nus will also contain any other new loved from elsewhere (Module 7b) the type species) that the gen	w species created and assigned to it (Module 2) and any that . Please enter here the TOTAL number of species aus will contain:	
1			

Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

Mavirus depends for its multiplication on the giant virus CroV, a member of the genus *Cafeteriavirus* within the family *Mimiviridae*. Both Mavirus and CroV infect heterotrophic nanoflagellates related to the marine protist *Cafeteria roenbergensis*; however, only co-infection with CroV leads to productive Mavirus infection. Mavirus contains a circular double-stranded DNA genome of 19,063 kbp that codes for 20 proteins (Module 10; Figure 1) [9]. The virions are icosahedral with a diameter of ~75 nm, and are composed of at least two different proteins with jelly-roll folds, the minor and the major capsid protein (Module 10; Figure 2a). In addition to the two capsid protein genes, members of the proposed genus encode a FtsK-HerA family DNA-packaging ATPase, a cysteine protease, a superfamily 3 helicase, a retroviral DDE-type integrase, a protein-primed family B DNA polymerase, and a Zinc-ribbon domain protein, as well as further proteins of unknown function [9, 16]. A distinct feature of Mavirus and related viruses is their striking genetic similarity to eukaryotic DNA transposons of the Maverick/Polinton family, with whom they share 7 homologous proteins [9, 11, 12, 17, 18]. Both, host-range and genomic content, distinguish viruses of the proposed genus from other members of the proposed family *Lavidaviridae* and justify the creation of a new genus.

Origin of the new genus name:

Ma- for Maverick. The Maverick/Polinton DNA transposons of eukaryotes are genetically related to the Mavirus.

Reasons to justify the choice of type species:

Mavirus is the first and sole isolated representative of this proposed genus and its genome sequence is available.

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

Membership in the genus is based on the dependence on a giant DNA virus of the genus *Cafeteriavirus* within the family *Mimiviridae*, and on the presence of a set of at least 7 homologous genes. These include a major and a minor capsid protein gene, a FtsK-HerA family DNA-packaging ATPase, a cysteine protease, a protein-primed family B DNA polymerase, a superfamily 3 helicase, and a retrovirus-type integrase [9, 16]. The capsid proteins of viruses in the proposed genus are homologous and the major capsid protein can be used as a phylogenetic marker to demonstrate membership in the proposed genus (Module 10; Figure 3). Species in the proposed genus display a close genetic relationship to eukaryotic DNA transposons of the Maverick/Polinton family [9, 11, 12, 18, 17].

MODUL	E 5: <u>NEW FAMILY</u>	
creating	g and naming a new family	
Code	2015.001iF	(assigned by ICTV officers)
To crea	ite a new family containing th	e subfamilies and/or genera listed below within the
Order:	unassigned	
If there is If the Or	s no Order, write " unassigned " he der has yet to be created (in Modu	re. le 6) please write " (new) " after the proposed name.
Code	2015.001jF	(assigned by ICTV officers)
To nam	ne the new family: Lavidavirida	ae
occionin	a subfamilies can are and unas	airmed appaires to a new family
assigni		signed species to a new ranning
Code		(assigned by ICTV officers)
•	If the subfamily is new, it must be o If the subfamily already exists, plea	reated in Module 4 ase complete Module 7 to 'REMOVE' it from its existing family
Code	2015.001kF	(assigned by ICTV officers)
To assig You may	gn the following genera to the y list several genera here. For each If the genus is new, it must be crea If the genus already exists, please from another family. If the latter, co	new family: a genus, please state whether it is new or existing. ated in Module 3 state whether it is currently unassigned or is to be removed omplete Module 7 to 'REMOVE' it from that family
Sputnik	<i>virus</i> (new)	
Maviru	s (new)	
The new	/ family will also contain any other i	new species created and assigned to it (Module 3) and any
unassig	med species that the family wi	ll contain (those NOT within any of the genera or
subfam	ilies listed above):	
0		
Reason Addition	s to justify the creation of the al material in support of this propos	new family: sal may be presented in the Appendix, Module 9
All viru	ses in the proposed family enco	de a conserved set of six proteins or domains, which
strongly	y suggests a monophyletic origin	1. This set consists of the morphogenetic module major
capsid p	protein (nexon protein), minor c	apsic protein (penton protein), FtsK-HerA family DNA- se as well as a primase-superfamily 3 belicase (S3H)
and a Z	inc-ribbon domain protein (Mod	dule 10, Figure 1) [17]. Although structurally
homolo	gous to jelly-roll capsid protein	s from other viruses, the capsid protein genes of these

viruses display no similarity to proteins outside this group with sequence-based homology detection methods such as BLASTp. Within the proposed family, however, the major capsid protein can be used as a phylogenetic marker to demonstrate membership with the family and to further subclassify members into genera (Module 10, Figure 3) [16, 19]. All viruses of the

proposed family, as well as further putative members from metagenomes without isolates, have circular or linear double-stranded DNA genomes that range in size from 17 kbp to 30 kbp and encode 16-34 ORFs [1, 9, 16, 20–22]. Cultured representatives have 40-80 nm icosahedral capsids with T=27 quasisymmetry (Module 10; Figure 1) [1, 2, 7, 8, 23]. All viruses in the proposed family depend on, or are found in association with, a large dsDNA virus of the family *Mimiviridae*. Co-infection with a suitable dsDNA virus is required for propagation of these viruses, as they replicate inside the cytoplasmic large virus factory and are presumed to use the transcription proteins encoded by the large dsDNA virus [1, 9, 24]. Viruses in the proposed family typically act as parasites of their associated large dsDNA viruses because they decrease the yield of the latter in co-infected cells, and have therefore been called "virophages" [1, 9, 14]. Based on these unifying characteristics which clearly set viruses of this group apart from other viruses, it seems appropriate to classify them within their own family.

Origin of the new family name:

La- for large, *vi*- for virus, *d*- for dependent, *a*- for associated (large virus dependent or associated)

MODULE 10: APPENDIX: supporting material

additional material in support of this proposal

References:

1.	La Scola B, Desnues C, Pagnier I, et al. (2008) The virophage as a unique parasite of the giant mimivirus. Nature 455:100–4. doi: 10.1038/nature07218
2.	Gaia M, Benamar S, Boughalmi M, et al. (2014) Zamilon, a novel virophage with mimiviridae host specificity. PLoS One 9:e94923. doi: 10.1371/journal.pone.0094923
3.	Arslan D, Legendre M, Seltzer V, et al. (2011) Distant Mimivirus relative with a larger genome highlights the fundamental features of Megaviridae. Proc Natl Acad Sci U S A 108:17486–17491. doi: 10.1073/pnas.1110889108
4.	Boughalmi M, Saadi H, Pagnier I, et al. (2013) High-throughput isolation of giant viruses of the Mimiviridae and Marseilleviridae families in the Tunisian environment. Env Microbiol 15:2000–7. doi: 10.1111/1462-2920.12068
5.	Desnues C, Boyer M, Raoult D (2012) Sputnik, a virophage infecting the viral domain of life. Adv Virus Res 82:63–89. doi: 10.1016/B978-0-12-394621-8.00013-3
6.	Desnues C, Raoult D (2010) Inside the Lifestyle of the Virophage. Intervirology 53:293–303.
7.	Sun S, La Scola B, Bowman VD, et al. (2010) Structural studies of the Sputnik virophage. J Virol 84:894–7. doi: 10.1128/JVI.01957-09
8.	Zhang X, Sun S, Xiang Y, et al. (2012) Structure of Sputnik, a virophage, at 3.5-A resolution. Proc Natl Acad Sci U S A 109:18431–6. doi: 10.1073/pnas.1211702109
9.	Fischer MG, Suttle CA (2011) A Virophage at the Origin of Large DNA Transposons. Science 332:231–234. doi: 10.1126/science.1199412
10.	Fischer MG, Allen MJ, Wilson WH, Suttle CA (2010) Giant virus with a remarkable complement of genes infects marine zooplankton. Proc Natl Acad Sci U S A 107:19508–13. doi: 10.1073/pnas.1007615107
11.	Pritham EJ, Putliwala T, Feschotte C (2007) Mavericks, a novel class of giant transposable elements widespread in eukaryotes and related to DNA viruses. Gene 390:3–17. doi: 10.1016/j.gene.2006.08.008
12.	Kapitonov V V, Jurka J (2006) Self-synthesizing DNA transposons in eukaryotes. Proc Natl Acad Sci U S A 103:4540–5. doi: 10.1073/pnas.0600833103
13.	Krupovic M, Bamford DH, Koonin E V (2014) Conservation of major and minor jelly- roll capsid proteins in Polinton (Maverick) transposons suggests that they are bona fide viruses. Biol Direct 9:6. doi: 10.1186/1745-6150-9-6
14.	Gaia M, Colson P, Desnues C, La Scola B (2013) The virophage concept. eLS 1–12.

additional material in support of this proposal

References:

 Desnues C, La Scola B, Yutin N, et al. (2012) Provirophages and transpovirons as the diverse mobilome of giant viruses. Proc Natl Acad Sci U S A 109:18078–83. doi: 10.1073/pnas.1208835109 Zhou J, Zhang W, Yan S, et al. (2013) Diversity of virophages in metagenomic data sets. J Virol 87:4225–36. doi: 10.1128/JVI.03398-12 Yutin N, Raoult D, Koonin E V (2013) Virophages, polintons, and transpovirons: a complex evolutionary network of diverse selfish genetic elements with different reproduction strategies. Virol J 10:158. doi: 10.1186/1743-422X-10-158 Krupovic M, Koonin E V. (2014) Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nat Rev Microbiol. doi: 10.1038/nrmicro3389 Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV-16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 		doi: 10.1002/9780470015902.a0024410
 Zhou J, Zhang W, Yan S, et al. (2013) Diversity of virophages in metagenomic data sets. J Virol 87:4225–36. doi: 10.1128/JVI.03398-12 Yutin N, Raoult D, Koonin E V (2013) Virophages, polintons, and transpovirons: a complex evolutionary network of diverse selfish genetic elements with different reproduction strategies. Virol J 10:158. doi: 10.1186/1743-422X-10-158 Krupovic M, Koonin E V. (2014) Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nat Rev Microbiol. doi: 10.1038/nrmicro3389 Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV-16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	15.	Desnues C, La Scola B, Yutin N, et al. (2012) Provirophages and transpovirons as the diverse mobilome of giant viruses. Proc Natl Acad Sci U S A 109:18078–83. doi: 10.1073/pnas.1208835109
 Yutin N, Raoult D, Koonin E V (2013) Virophages, polintons, and transpovirons: a complex evolutionary network of diverse selfish genetic elements with different reproduction strategies. Virol J 10:158. doi: 10.1186/1743-422X-10-158 Krupovic M, Koonin E V. (2014) Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nat Rev Microbiol. doi: 10.1038/nrmicro3389 Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV-16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	16.	Zhou J, Zhang W, Yan S, et al. (2013) Diversity of virophages in metagenomic data sets. J Virol 87:4225–36. doi: 10.1128/JVI.03398-12
 Krupovic M, Koonin E V. (2014) Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nat Rev Microbiol. doi: 10.1038/nrmicro3389 Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	17.	Yutin N, Raoult D, Koonin E V (2013) Virophages, polintons, and transpovirons: a complex evolutionary network of diverse selfish genetic elements with different reproduction strategies. Virol J 10:158. doi: 10.1186/1743-422X-10-158
 Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	18.	Krupovic M, Koonin E V. (2014) Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nat Rev Microbiol. doi: 10.1038/nrmicro3389
 Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	19.	Yutin N, Kapitonov V V, Koonin E V (2015) A new family of hybrid virophages from an animal gut metagenome. Biol Direct 10:1–9. doi: 10.1186/s13062-015-0054-9
 Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14 Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110 	20.	Yau S, Lauro FM, DeMaere MZ, et al. (2011) Virophage control of antarctic algal host-virus dynamics. Proc Natl Acad Sci USA 108:6163–8. doi: 10.1073/pnas.1018221108
22. Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110	21.	Zhou J, Sun D, Childers A, et al. (2015) Three Novel Virophage Genomes Discovered from Yellowstone Lake. J Virol 89:1278–1285. doi: 10.1128/JVI.03039-14
	22.	Santini S, Jeudy S, Bartoli J, et al. (2013) Genome of Phaeocystis globosa virus PgV- 16T highlights the common ancestry of the largest known DNA viruses infecting eukaryotes. Proc Natl Acad Sci U S A. doi: 10.1073/pnas.1303251110
 Campos RK, Boratto P V, Assis FL, et al. (2014) Samba virus: a novel mimivirus from a giant rain forest, the Brazilian Amazon. Virol J 11:95. doi: 10.1186/1743-422X-11-95 	23.	Campos RK, Boratto P V, Assis FL, et al. (2014) Samba virus: a novel mimivirus from a giant rain forest, the Brazilian Amazon. Virol J 11:95. doi: 10.1186/1743-422X-11-95
24. Claverie J-M, Abergel C (2009) Mimivirus and its virophage. Annu Rev Genet 43:49– 66. doi: 10.1146/annurev-genet-102108-134255	24.	Claverie J-M, Abergel C (2009) Mimivirus and its virophage. Annu Rev Genet 43:49– 66. doi: 10.1146/annurev-genet-102108-134255
25. Krupovič M, Bamford DH (2010) Order to the viral universe. J Virol 84:12476–9. doi: 10.1128/JVI.01489-10	25.	Krupovič M, Bamford DH (2010) Order to the viral universe. J Virol 84:12476–9. doi: 10.1128/JVI.01489-10
26. Raoult D, Forterre P (2008) Redefining viruses: lessons from Mimivirus. Nat Rev Microbiol 6:315–319. doi: 10.1038/nrmicro1858	26.	Raoult D, Forterre P (2008) Redefining viruses: lessons from Mimivirus. Nat Rev Microbiol 6:315–319. doi: 10.1038/nrmicro1858
27. King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (2011) Virus Taxonomy. Ninth Report of the International Committee on Taxonomy of Viruses. Elsevier Academic, London	27.	King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (2011) Virus Taxonomy. Ninth Report of the International Committee on Taxonomy of Viruses. Elsevier Academic, London
28. Krupovic M, Cvirkaite-Krupovic V (2011) Virophages or satellite viruses? Nat Rev	28.	Krupovic M, Cvirkaite-Krupovic V (2011) Virophages or satellite viruses? Nat Rev

additional material in support of this proposal

References:





Figure 1. Comparative genomic maps of the virophages OLV (Organic Lake virophage), Sputnik, and Mavirus. ORFs are indicated with arrows. Conserved virophage genes are shown in colour: Superfamily 3 helicase, pink; Zinc-ribbon domain, yellow; FtsK-HerA family ATPase, red; Cys protease, green; minor capsid protein, light blue; major capsid protein, indigo. The scale bar shows distances in kilobase pairs.



Figure 2. The virions of Mavirus and Sputnik. A. Cryo-EM reconstruction of the Sputnik virion (adapted from Ref. [8], Electron Microscopy Data Bank ID 5495). B. Negative stain electron micrograph of Mavirus particles (U. Mersdorf, Max Planck Institute for Medical Research).



Figure 3. Phylogenetic analysis of the major capsid proteins of virophages. Branches are coloured according to the proposed genera of virophages: "*Sputnikvirus*", blue; "*Mavirus*", red. The proposed type species of the two tentative genera are designated with asterisks. ALM represents a metagenomic virophage, whose genome sequence makes it a member of the proposed genus *Mavirus*. However, since no isolates for ALM are available, it cannot be classified at this point. The multiple sequence alignment for phylogenetic analysis was constructed using PROMALS3D

[9] with the Sputnik Cryo-EM structure (protein data bank ID 3j26) as 3D structure template. Columns containing gaps were removed from the alignment. Maximum-likelihood phylogenetic analysis was carried out using PhyML 3.1 [10], with the Whelan and Goldman (WAG) model of amino acid substitutions, including a gamma law with four substitution rate categories. Numbers at the branch points represent SH (Shimodaira–Hasegawa)-like local support values. The scale bar represents the number of substitutions per site. All taxa are indicated with the corresponding GenBank identifiers, or in the case of rumen virophages with the Shotgun Assembly Sequence identifier. Abbreviations: ALM, Ace Lake mavirus; OLV, Organic Lake virophage; RVP, rumen virophage; YSLV, Yellowstone Lake virophage.