

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2016.056	(to be completed by ICTV officers)			
Short title: To create one (1) family Siphoviridae. (e.g. 6 new species in the genus Modules attached (modules 1 and 10 are required)		ponusvirus, including one (1) new species in the $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			
Author(s):					
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List the ICTV study group(s) that have seen this proposal:					
A list of study groups and contact http://www.ictvonline.org/subcom in doubt, contact the appropriate chair (fungal, invertebrate, plant, vertebrate viruses)	mittees.asp . If subcommittee	ICTV Bacterial and Archaeal Viruses Subcommittee			
ICTV Study Group comments (if any) and response of the proposer:					
Date first submitted to ICTV: Date of this revision (if different					
ICTV-EC comments and response of the proposer:					

MODULE 2: NEW SPECIES

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	201	6.056aB	(assigned by IC			
To crea	te 1 ne	ew species within	:			
G	enus:	Cranusvirus (no	aw)		that apply.	
	mily:			created (in a later module, below) write "(new)" after its proposed name. • If no genus is specified, enter		
Fa	mily:	Siphoviridae				
(Order:	Caudovirales		"unassigned" in the genus box.		
Name of new species:		species:	Representative isolate: (only 1 per species please)		GenBank sequence accession number(s)	
Rhodobacter virus RcCronus		irus RcCronus	Rhodobacter phage R	cCronus	KR935217	

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, explain how the new species meet these criteria.
 - o If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 3: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	201	6.056bB	(assigned by ICTV officers)		
To create	a new	genus within:		Fill in all that apply.	
Subfa	mily:			If the higher taxon has yet to be created	
Fai	mily:	Siphoviridae		(in a later module, below) write "(new)" after its proposed name.	
O	rder:	Caudovirales		 If no family is specified, enter "unassigned" in the family box 	

naming a new genus

Code	2016.056cB	(assigned by ICTV officers)
To name the new genus: Cronusvirus		

Assigning the type species and other species to a new genus

Assigning the type species and other species to a new genus					
Code	2016.056dB	(assigned by ICTV officers)			
To designa	To designate the following as the type species of the new genus				
Rhodobaci	Rhodobacter virus RcCronus Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered				
The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). Please enter here the TOTAL number of species (including the type species) that the genus will contain: 1					

Reasons to justify the creation of a new genus:

Additional material in support of this proposal may be presented in the Appendix, Module 9

The newly isolated bacteriophages for this proposed genus, Rhodobacter phage RcCronus and its strains Rhodobacter phages RcRhea and RcSaxon, were isolated from stream water collected in McLean County, Illinois, USA. These phages were isolated on *Rhodobacter capsulatus* as a host and are distinct from other bacteriophages that have been isolated using this host. "The genomes have defined ends with 13 base 5' extensions. Notable features include a –1 translational frameshift for a tail assembly protein, a DNA methylase, a GTA-related tail protein, RepA, and a plasmid partitioning protein.[4]" At present there is no electron micrograph of this virus. The genomes of phages belonging to this genus show no sequence identity to any other phage in the NCBI database.

BLASTN and phylogenetic analyses (Fig. 2) [3] all indicate that the proposed genus, *Cronusvirus*, is cohesive and distinct from other genera. On average, the genomes of members of this genus are 36.0 kb in length (65.4 mol% G+C), and encode 45 proteins and 0 tRNAs.

Origin of the new genus name:

Based upon the name of the second sequenced member of this genus.

Reasons to justify the choice of type species:

Usually this genus would be called after the first sequence phage RcRhea, but Rheavirus sounds too similar to *Reovirus*.

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 10: **APPENDIX**: supporting material

additional material in support of this proposal

References:

- 1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.
- 2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.
- 3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.
- 4. Bollivar DW, Bernardoni B, Bockman MR, Miller BM, Russell DA, Delesalle VA, Krukonis GP, Hatfull GF, Cross MR, Szewczyk MM, Eppurath A. Complete Genome Sequences of Five Bacteriophages That Infect *Rhodobacter capsulatus*. Genome Announc. 2016 May 26;4(3). pii: e00051-16.

Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

Table 1. Properties of Rhodobacter phage RcCronus.

Rhodobacter phage	GenBank Accession No.	Genome length (kb)	Genome (mol% G+C)	No. CDS
RcCronus	KR935217	35.98	65.4	44

Rhodobacter phages RcRhea (KR935216) and RcSaxon (KT253150) should be considered as strains of phage RcCronus.

Fig. 2. Phylogenetic analysis of (A) large subunit terminase proteins, and (B) major capsid proteins of cronusviruses and homologous proteins from a variety of other phages constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."

A. TerL protein

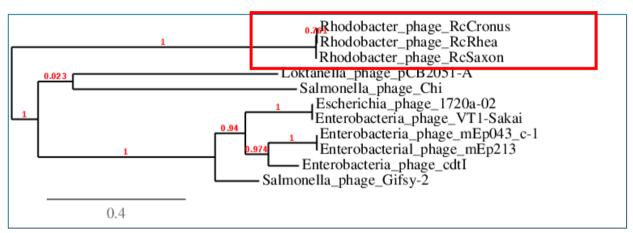


Figure 1: Phylogenetic tree (the branch length is proportional to the number of substitutions per site).

B. Major capsid protein

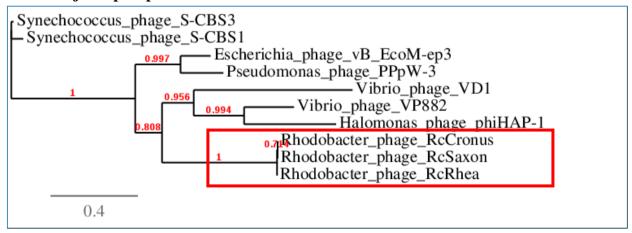


Figure 1: Phylogenetic tree (the branch length is proportional to the number of substitutions per site).