

This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

# MODULE 1: TITLE, AUTHORS, etc

Code assigned:	2016.021a-a	lB	(to be completed by ICTV officers)		
Short title: To create one (1) r Podoviridae. (e.g. 6 new species in the genus A Modules attached (modules 1 and 10 are required)		$\approx$ , including one $\boxed{2}$	e(1) new s $3 \boxtimes$	species in ↑ 4 □ 9 □	5
	0		δ	9	10 🖂

Author(s):

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#### List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided at <u>http://www.ictvonline.org/subcommittees.asp</u> . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses)	ICTV Subcom	Bacterial nmittee	and	Archaeal	Viruses
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### ICTV Study Group comments (if any) and response of the proposer:

Date first submitted to ICTV: Date of this revision (if different to above): June 2016

#### **ICTV-EC** comments and response of the proposer:

# MODULE 2: NEW SPECIES

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code 201	Code 2016.021aB (assigned by IC			CTV officers)			
To create 1 n	To create 1 new species within:						
				all that apply.			
Genus: <i>Kf1virus</i> (new)			If the higher taxon has yet to be				
Subfamily:			<ul> <li>created (in a later module, below) write</li> <li>"(new)" after its proposed name.</li> <li>If no genus is specified, enter</li> </ul>				
Family:	Podoviridae						
Order:	Order: Caudovirales			"unassigned" in the genus box.			
		Representative isolate 1 per species please)	e: (only	GenBank sequence accession number(s)			
Edwardsiella virus KF1 Edward		Edwardsiella phage KI	F-1	AB757800			

**Reasons to justify the creation and assignment of the new species:** 

- Explain how the proposed species differ(s) from all existing species.
  - If species demarcation criteria (see module 3) have previously been defined for the genus, explain how the new species meet these criteria.
    - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

## MODULE 3: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	201	6.021bB	(assigned by IC	(assigned by ICTV officers)				
To create	a new	genus within:		Fill in all that apply.				
Subfai	mily:			• If the higher taxon has yet to be created				
Fai	mily:	Podoviridae		(in a later module, below) write "( <b>new)</b> " after its proposed name.				
0	order:	Caudovirales		<ul> <li>If no family is specified, enter</li> <li>"unassigned" in the family box</li> </ul>				

naming a new genus

Code	2016.021cB	(assigned by ICTV officers)
To name tl	ne new genus: <i>Kf1virus</i>	

Assigning the type species and other species to a new genus

Code	2016.021dB	(assigned by ICTV officers)					
To designa	To designate the following as the type species of the new genus						
Edwardsiella virus KF1		Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered					
The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). <b>Please enter here the TOTAL number of species</b>							

are being moved from elsewhere (Module 7b). Please enter here the TOTAL number of species (including the type species) that the genus will contain:

1

### **Reasons to justify the creation of a new genus:**

Additional material in support of this proposal may be presented in the Appendix, Module 9

*Edwardsiella tarda* lytic bacteriophages KF-1 and IW-1 are members of the *Podoviridae* [4]. They were isolated from Japanese flounder kidney (Edwardsiella phage KF-1) and seawater (Edwardsiella phage IW-1).

BLASTN, CoreGenes (Table 1) [2], and phylogenetic analyses (Fig. 1) [3] all indicate that the proposed genus, *Kflvirus*, is cohesive and distinct from other genera. On average the genomes of members of this genus are 41.6 kb in length (48.3 mol% G+C), and encode 48 proteins and 0 tRNAs.

### **Origin of the new genus name:**

Based upon the name of the first sequenced member of this genus.

### **Reasons to justify the choice of type species:**

The first sequenced member of this genus.

#### Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

# MODULE 10: APPENDIX: supporting material

additional material in support of this proposal

### **References:**

1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.

2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.

3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.

4. Yasuike M, Sugaya E, Nakamura Y, Shigenobu Y, Kawato Y, Kai W, Fujiwara A, Sano M, Kobayashi T, Nakai T. Complete Genome Sequences of *Edwardsiella tarda*-Lytic Bacteriophages KF-1 and IW-1. Genome Announc. 2013;1(1). pii: e00089-12.

### Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

**Table 1**. Properties of the two phages belonging to the genus *Kflvirus*.

Edwardsi ella phage	RefSeq No.	GenBank Accession No.	Genome length (kb)	Genome (mol% G+C)	No. CDS	DNA (% sequence identity)*	% Homolo gous proteins **
KF-1	NC_019420.1	AB757800.1	41.55	48.3	48	100	100
IW-1***		AB757801.1	41.68	48.3	48	99	97.9

\* Determined using BLASTN; \*\* Determined using CoreGenes [2]; \*\*\* a strain of Edwardsiella phage KF-1.

**Fig. 1.** Phylogenetic analysis of large subunit terminase proteins of kf1viruses and a variety of other homologous phage proteins constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."

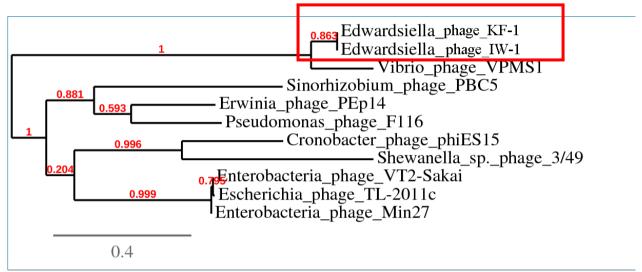


Figure 1: Phylogenetic tree (the branch length is proportional to the number of substitutions per site).