MODULE 1: **TITLE, AUTHORS, etc**

Code assigned:	*2016.001a-dB*	(to be completed by ICTV officers)

Short title: To create one (1) new genus, *Ab18virus*, including three (3) new species in the family *Siphoviridae*.
(e.g. 6 new species in the genus *Zetavirus*)

Modules attached
(modules 1 and 10 are required)

1 ☒	2 ☒	3 ☒	4 ☐	5 ☐
6 ☐	7 ☐	8 ☐	9 ☐	10 ☒

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List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided at http://www.ictvonline.org/subcommittees.asp . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses)	ICTV Bacterial and Archaeal Viruses Subcommittee

ICTV Study Group comments (if any) and response of the proposer:

Date first submitted to ICTV: June, 2016
Date of this revision (if different to above):

ICTV-EC comments and response of the proposer:

MODULE 2: **NEW SPECIES**

creating and naming one or more new species.
If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be "unassigned" within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	*2016.001aB*		(assigned by ICTV officers)

To create 3 new species within:

		Fill in all that apply.
Genus:	*Ab18virus* **(new)**	• If the higher taxon has yet to be created (in a later module, below) write "**(new)**" after its proposed name.
Subfamily:		
Family:	*Siphoviridae*	• If no genus is specified, enter "**unassigned**" in the genus box.
Order:	*Caudovirales*	

Name of new species:	Representative isolate: (only 1 per species please)	GenBank sequence accession number(s)
Pseudomonas virus Ab18	Pseudomonas phage vB_PaeS_PAO1_Ab18	LN610577
Pseudomonas virus Ab19	Pseudomonas phage vB_PaeS_PAO1_Ab19	LN610584
Pseudomonas virus PaMx11	Pseudomonas phage PaMx11	JQ067087

Reasons to justify the creation and assignment of the new species:
- Explain how the proposed species differ(s) from all existing species.
  - If species demarcation criteria (see module 3) have previously been defined for the genus, **explain how the new species meet these criteria**.
  - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 9

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 3: **NEW GENUS**

creating a new genus
Ideally, a genus should be placed within a higher taxon.

Code	*2016.001bB*	(assigned by ICTV officers)

To create a new genus within:

Fill in all that apply.

Subfamily:	
Family:	*Siphoviridae*
Order:	*Caudovirales*

- If the higher taxon has yet to be created (in a later module, below) write "**(new)**" after its proposed name.
- If no family is specified, enter "**unassigned**" in the family box

naming a new genus

Code	*2016.001cB*	(assigned by ICTV officers)

To name the new genus: *Ab18virus*

Assigning the type species and other species to a new genus

Code	*2016.001dB*	(assigned by ICTV officers)

To designate the following as the type species of the new genus

Pseudomonas virus Ab18

Every genus must have a type species. This should be a well characterized species although not necessarily the first to be discovered

The new genus will also contain any other new species created and assigned to it (Module 2) and any that are being moved from elsewhere (Module 7b). **Please enter here the TOTAL number of species (including the type species) that the genus will contain:**

3

Reasons to justify the creation of a new genus:
Additional material in support of this proposal may be presented in the Appendix, Module 9

These *Pseudomonas* phages were isolated from Côte d'Ivoire [4] and Mexico [5].
BLASTN, CoreGenes (Table 1) [2], progressiveMauve alignment (Fig. 2) [1] and phylogenetic analyses (Fig. 3) [3] all indicate that the proposed genus, *Ab18virus*, is cohesive and distinct from other genera. On average the genomes of members of this genus are 58.2kb (63.7 mol% G+C), and encode 78 proteins and 0 tRNAs.
Modification of the genome protects it against digestion by several restriction enzymes, although the sites are present.
The genome of Ab18 possesses a repressor and an integrase but no lysogens could be obtained from PAO1 and another clinical strain on which Ab18 produces turbid plaques.

Origin of the new genus name:

Based upon the name of the first sequenced member of this genus

Reasons to justify the choice of type species:

The first sequenced member of this genus

Species demarcation criteria in the new genus:

We have chosen 95% DNA sequence identity as the criterion for demarcation of species in this new genus. The members of each of the proposed species differ from those of other species by more than 5% at the DNA level as confirmed with the BLASTN algorithm.

MODULE 10: **APPENDIX**: supporting material

additional material in support of this proposal

References:

1. Darling AE, Mau B, Perna NT. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One. 2010; 5(6):e11147.

2. Turner D, Reynolds D, Seto D, Mahadevan P. CoreGenes3.5: a webserver for the determination of core genes from sets of viral and small bacterial genomes. BMC Res Notes. 2013; 6:140. doi: 10.1186/1756-0500-6-140.

3. Dereeper A, Guignon V, Blanc G, Audic S, Buffet S, Chevenet F, Dufayard JF, Guindon S, Lefort V, Lescot M, Claverie JM, Gascuel O. Phylogeny.fr: robust phylogenetic analysis for the non-specialist. Nucleic Acids Res. 2008; 36(Web Server issue):W465-9.

4. Essoh C, Latino L, Midoux C, Blouin Y, Loukou G, Nguetta SP, Lathro S, Cablanmian A, Kouassi AK, Vergnaud G, Pourcel C. Investigation of a Large Collection of *Pseudomonas aeruginosa* Bacteriophages Collected from a Single Environmental Source in Abidjan, Côte d'Ivoire. PLoS One. 2015; 10(6):e0130548.

5. Sepúlveda-Robles O, Kameyama L, Guarneros G. High diversity and novel species of *Pseudomonas aeruginosa* bacteriophages. Appl Environ Microbiol. 2012;78(12):4510-5.

Annex:

Fig. 1. Electron micrograph of negatively stained (2% potassium phosphotungstate (pH 7.0)) Pseudomonas phage Ab18 .
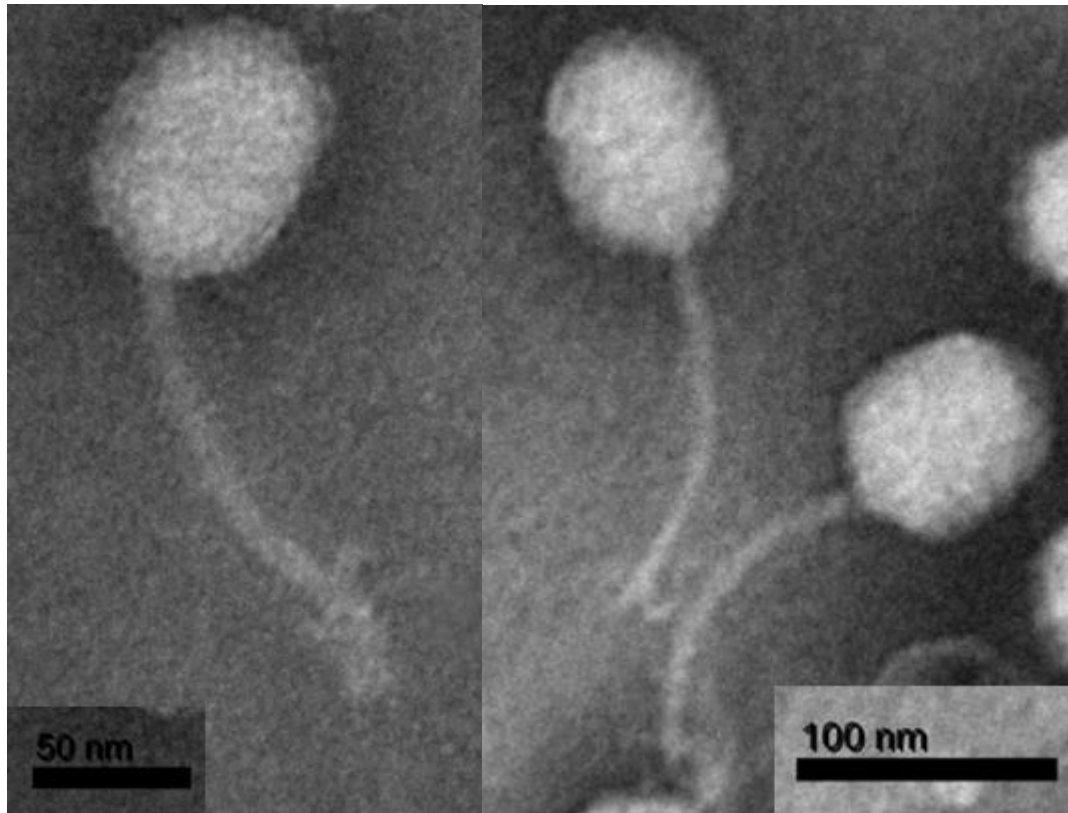
Table 1. Properties of the three phages belonging to the genus *Ab18virus*.

Phage	RefSeq No.	GenBank Accession No.	Genome size (kb)	Genome (mol%G+C)	No. CDS	DNA (% sequence identity)*	% Homologous proteins **
Ab18	NC_026594.1	LN610577.1	56.54	63.5	76	100	100
Ab19		LN610584.1	58.14	63.3	***	91	***
PaMx11		JQ067087.1	59.88	64.4	81	85	96.0

⁎ Determined using BLASTN; ** Determined using CoreGenes [2]; *** not annotated (81, Pourcel, personal communication); Pseudomonas phage vB_PaeS_PAO1_Ab20 (LN610585) should be considered as strain of Pseudomonas phage vB_PaeS_PAO1_Ab18 within this genus.

Fig. 2. Progressive Mauve alignment [1] of the annotated genomes of members of the *Ab18virus* genus – from top to bottom: PaMx11, Ab18, and Ab19. Colored blocks indicate the regions of 1 to 1 best alignment with rearrangement breakpoints in a different random color. The degree of sequence similarity between regions is given by a similarity plot within the colored blocks with the height of the plot proportional to the average nucleotide identity (Aaron Darling, personal communication). N.B. The first genome is not collinear with the next two.
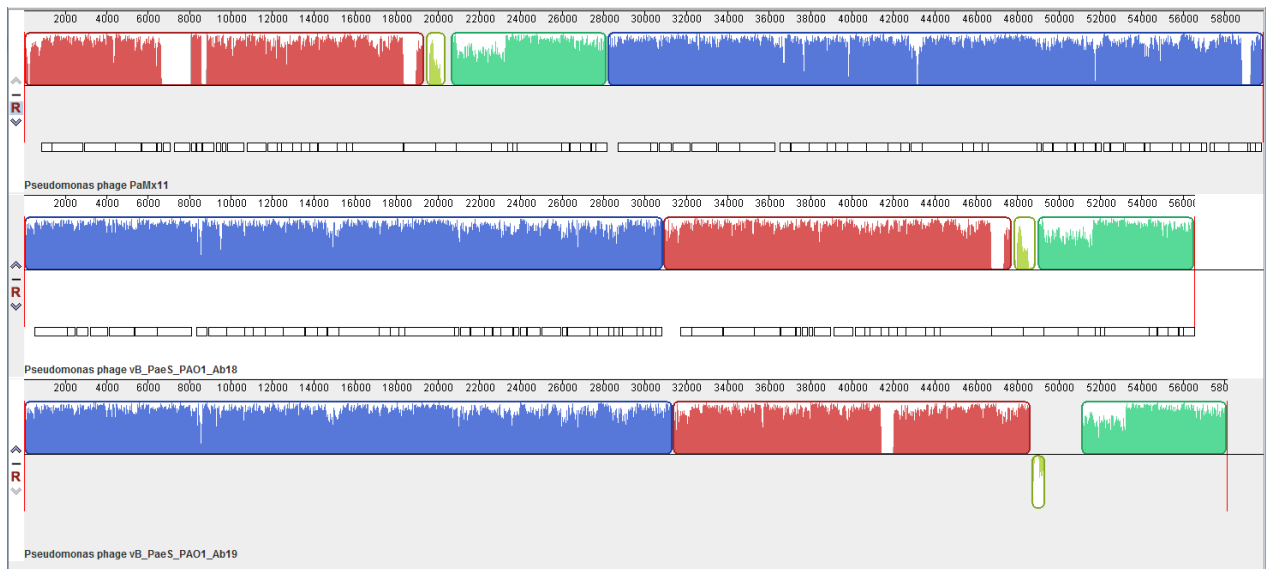
Fig. 3. Phylogenetic analysis of large subunit terminase proteins of ab18viruses and homologous proteins from a variety of other phages constructed using "one click" at phylogeny.fr [3]. "The "One Click mode" targets users that do not wish to deal with program and parameter selection. By default, the pipeline is already set up to run and connect programs recognized for their accuracy and speed (MUSCLE for multiple alignment and PhyML for phylogeny) to reconstruct a robust phylogenetic tree from a set of sequences." It also includes the use of Gblocks to eliminate poorly aligned positions and divergent regions. "The usual bootstrapping procedure is replaced by a new confidence index that is much faster to compute. See: Anisimova M., Gascuel O. Approximate likelihood ratio test for branches: A fast, accurate and powerful alternative (Syst Biol. 2006;55(4):539-52.) for details."
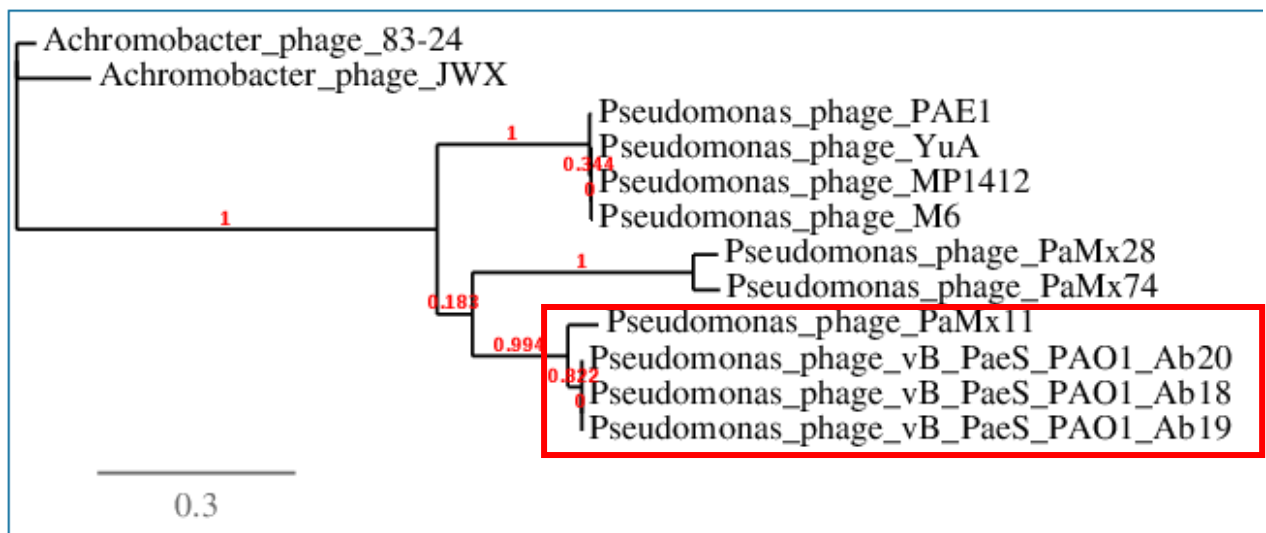

Figure 1: *Phylogenetic tree (the branch length is proportional to the number of substitutions per site).*