



This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

MODULE 1: **TITLE, AUTHORS, etc**

Code assigned:	2011.010a-eP	(to be completed by ICTV officers)			
Short title: create new genus, <i>Gallantivirus</i> , in the family <i>Tombusviridae</i>					
Modules attached	1 <input checked="" type="checkbox"/>	2 <input checked="" type="checkbox"/>	3 <input checked="" type="checkbox"/>	4 <input type="checkbox"/>	5 <input type="checkbox"/>
	6 <input type="checkbox"/>	7 <input checked="" type="checkbox"/>	8 <input type="checkbox"/>	9 <input checked="" type="checkbox"/>	

Author(s) with e-mail address(es) of the proposer:

D'Ann Rochon (dann.rochon@agr.gc.ca). On behalf of the Tombusviridae SG

List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided at <http://www.ictvonline.org/subcommittees.asp> . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses)

Tombusviridae

ICTV-EC or Study Group comments and response of the proposer:

Date first submitted to ICTV:

8 August 2011

Date of this revision (if different to above):

June 26 2012

1MODULE 3: NEW GENUS

creating a new genus

Ideally, a genus should be placed within a higher taxon.

Code	2011.010aP	(assigned by ICTV officers)
To create a new genus within:		
Subfamily:		
Family:	Tombusviridae	
Order:		

naming a new genus

Code	2011.010bP	(assigned by ICTV officers)
To name the new genus: <i>Gallantivirus</i>		

Assigning the type species and other species to a new genus

Code	2011.010cP	(assigned by ICTV officers)
To designate the following as the type species of the new genus		
<i>Galinsoga mosaic virus</i>		
Please enter here the TOTAL number of species (including the type species) that the genus will contain:		
1		

Reasons to justify the creation of a new genus:

[Additional material in support of this proposal may be presented in the Appendix, Module 9](#)

Genera in the family *Tombusviridae* are distinguished by genome organization and by the properties of the replication protein and the coat protein. Necroviruses have a distinctive type of coat protein without a protruding domain and forming a single lineage but their replication proteins are of two different lineages. There is therefore a current proposal to split the genus *Necrovirus* into two genera, *Alphanecrovirus* and *Betanecrovirus* (2011.009a-mP).

The complete sequence of *Galinsoga mosaic virus* (GaMV) shows that this virus is similar to alphanecroviruses in genome organization and in percent sequence identity in the replication protein and the two small movement protein ORFs (Tables 1-4). This is confirmed by phylogenetic analyses (Figs 3 and 4).

However, unlike necroviruses, the coat protein of GaMV contains a protruding domain (Fig. 1, 2) and it is most similar to the coat proteins of other members of the family with protruding domains in both sequence comparisons (Table 4) and phylogenetic analyses (Fig. 5).

To be consistent with genus demarcation criteria being applied in the family, a new genus is therefore needed to contain GaMV.

Origin of the new genus name:

Derived from "Gallant soldier", the vernacular English name of *Galinsoga parviflora*, the host of the type species of the genus

Reasons to justify the choice of type species:

This is a monotypic genus

Species demarcation criteria in the new genus:

If there will be more than one species in the new genus, list the criteria being used for species demarcation and explain how the proposed members meet these criteria.

< 80% amino acid sequence identity in the polymerase;
<55% amino acid sequence similarity in the coat protein.

MODULE 7: **REMOVE and MOVE**

Use this module whenever an existing taxon needs to be removed:

- Either to abolish a taxon entirely (when only part (a) needs to be completed)
- Or to move a taxon and re-assign it e.g. when a species is moved from one genus to another (when BOTH parts (a) and (b) should be completed)

Part (a) taxon/taxa to be removed or moved

Code	2011.010dP	(assigned by ICTV officers)
To remove the following taxon (or taxa) from their present position:		
<i>Galinsoga mosaic virus</i>		
The present taxonomic position of these taxon/taxa:		
Genus:	<i>Carmovirus</i>	Fill in all that apply.
Subfamily:		
Family:	<i>Tombusviridae</i>	
Order:		
If the taxon/taxa are to be abolished (i.e. not reassigned to another taxon) write "yes" in the box on the right		

Reasons to justify the removal:

Virus is being proposed to represent a new genus (this proposal)

Part (b) re-assign to a higher taxon

Code	2011.010a-eP	(assigned by ICTV officers)
To re-assign the taxon (or taxa) listed in Part (a) as follows:		
Genus:	<i>Gallantivirus</i> (new genus)	
Subfamily:		
Family:	<i>Tombusviridae</i>	
Order:		

Reasons to justify the re-assignment:

See Modules 3 and 9

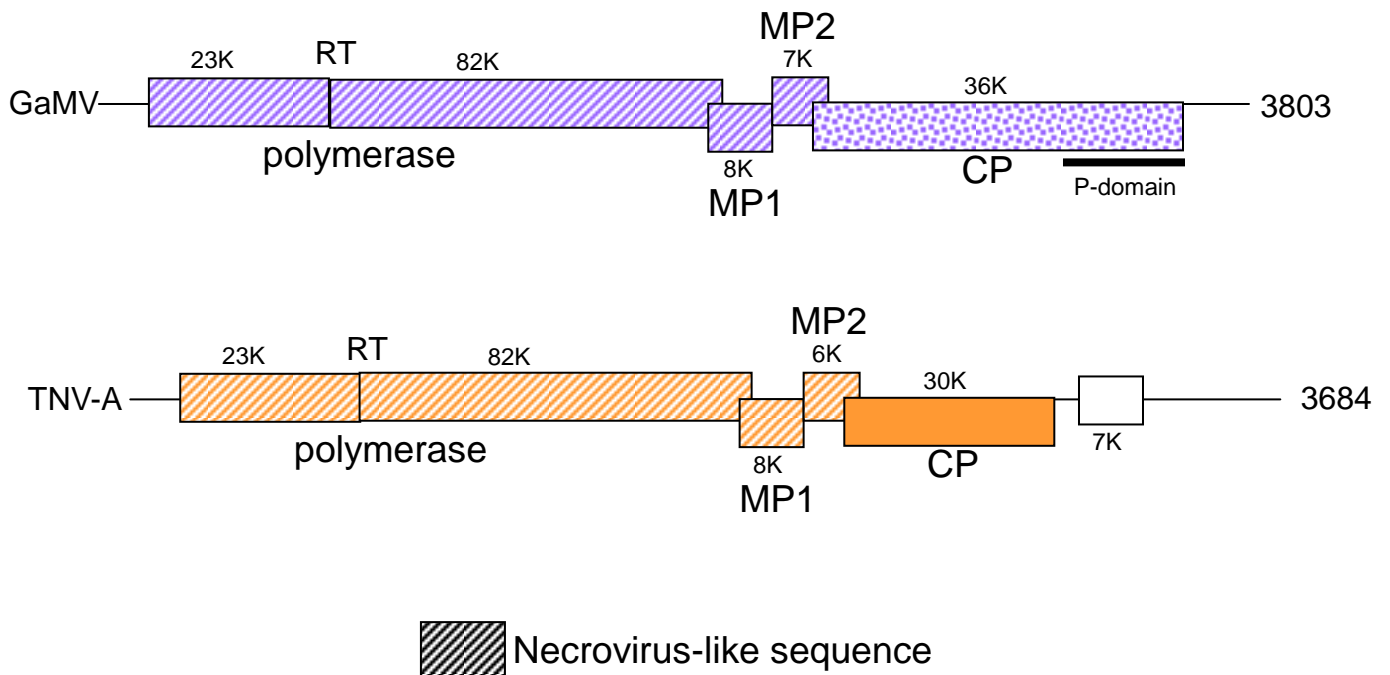
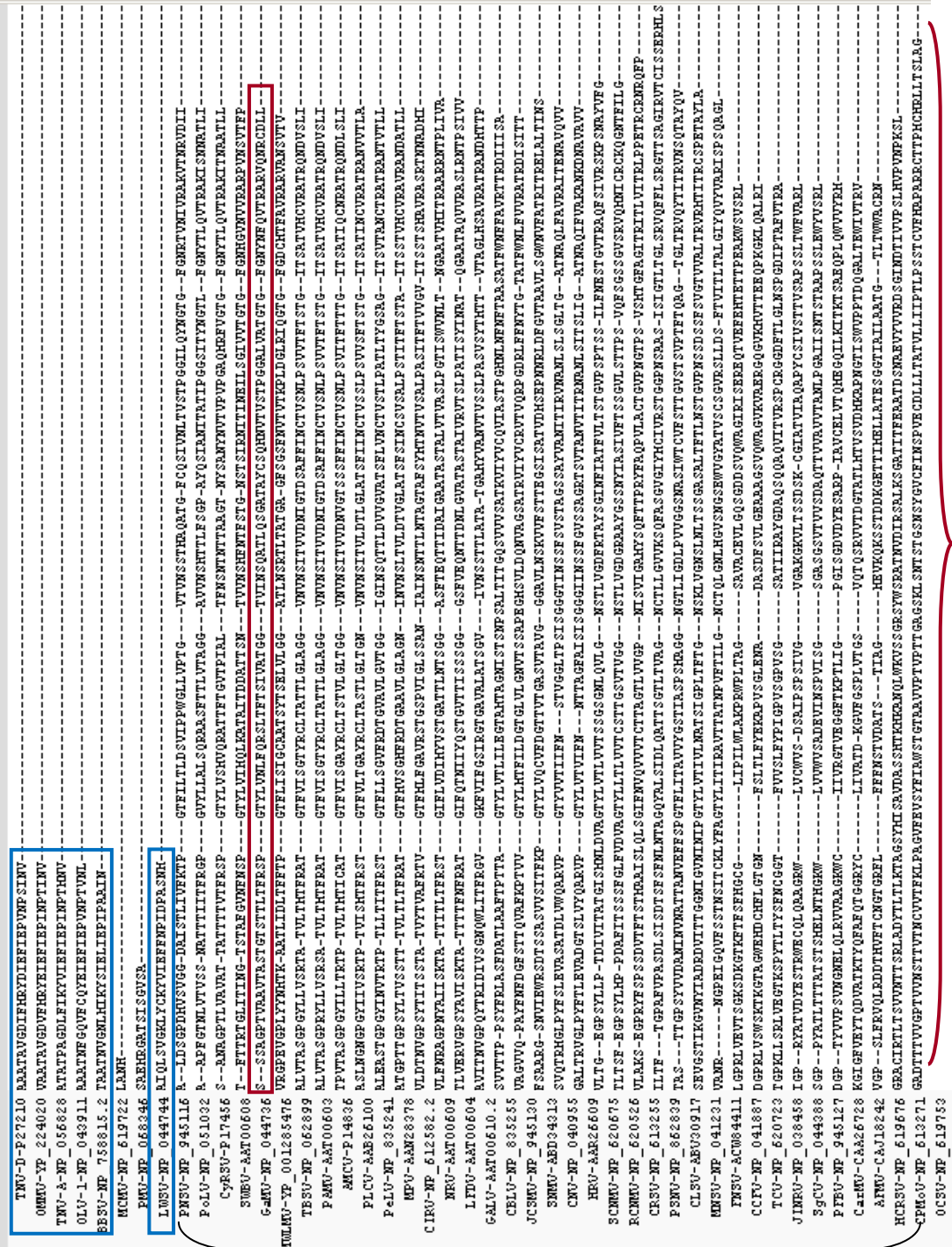


Figure 1. Genome organization of GaMV and comparison to tobacco necrosis virus A (TNV-A), the type member of the newly proposed genus *Alphanecrovirus*. Similar shading patterns indicate significant sequence identity. Note that the coat protein (CP) of GaMV, unlike TNV-A, contains a C-terminal protruding domain which is typical of the currently classified viruses in the *Tombusviridae* with the exception of the necroviruses.



Necrovirus

Other Tombusviridae members (with P domain)

Figure 2. Alignment of the CP sequence of GaMV (boxed in red) with the CPs of the proposed alpha and betanecroviruses (boxed in blue) and remaining members of the *Tombusviridae*. Only the C-terminal region of the CP alignment is shown with the area corresponding to the P-domain being shown in brackets.

Coat protein protrusion

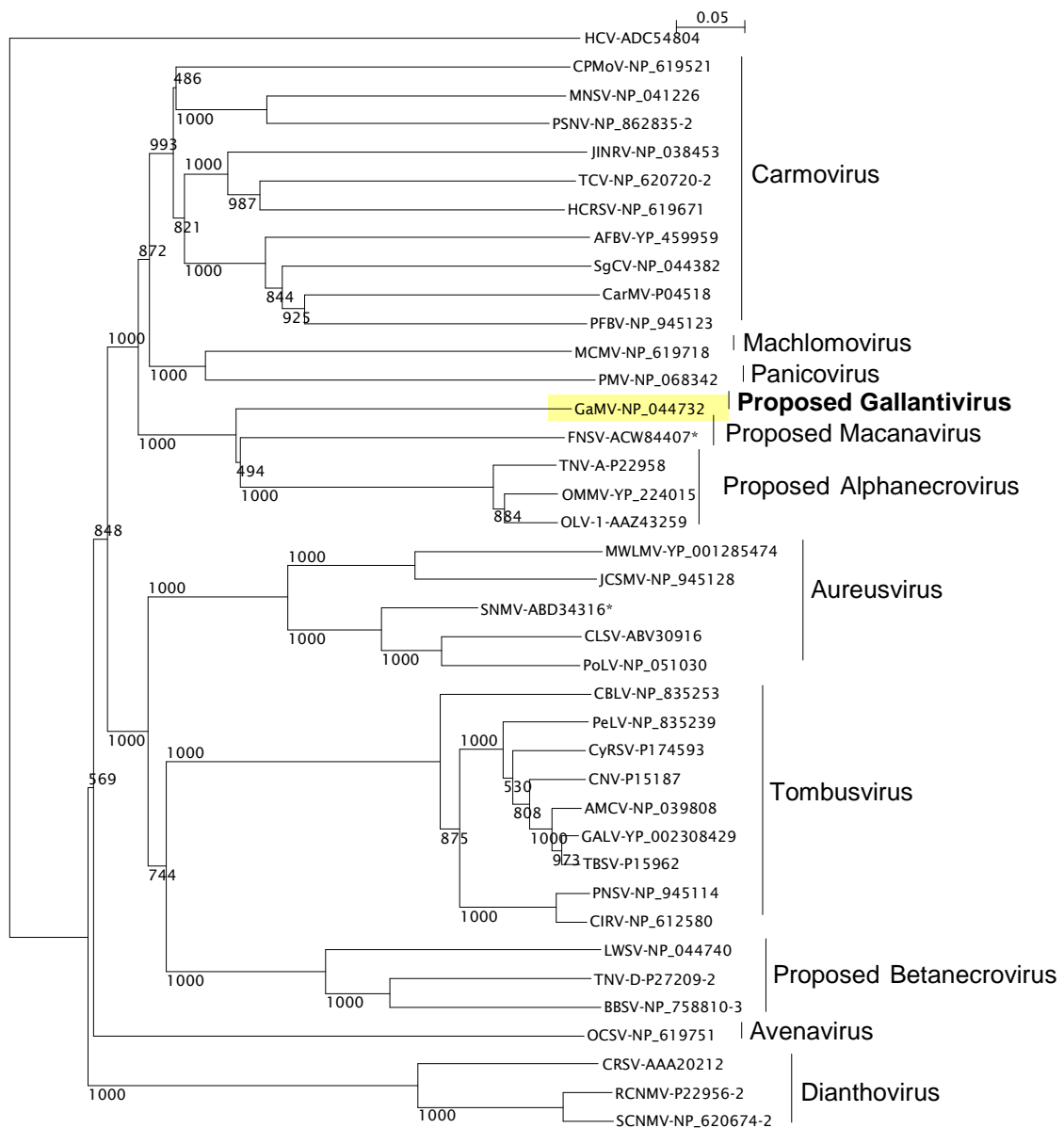


Figure 3. Phylogenetic (distance) tree of the polymerase of *Tombusviridae* members with the position of GaMV highlighted in yellow. Sequences were aligned using the ClustalX 2.1 algorithm and trees were generated by the Neighbor Joining method using 1000 bootstrap replicates. Hepatitis C virus (HCV) polymerase was used as an outgroup.

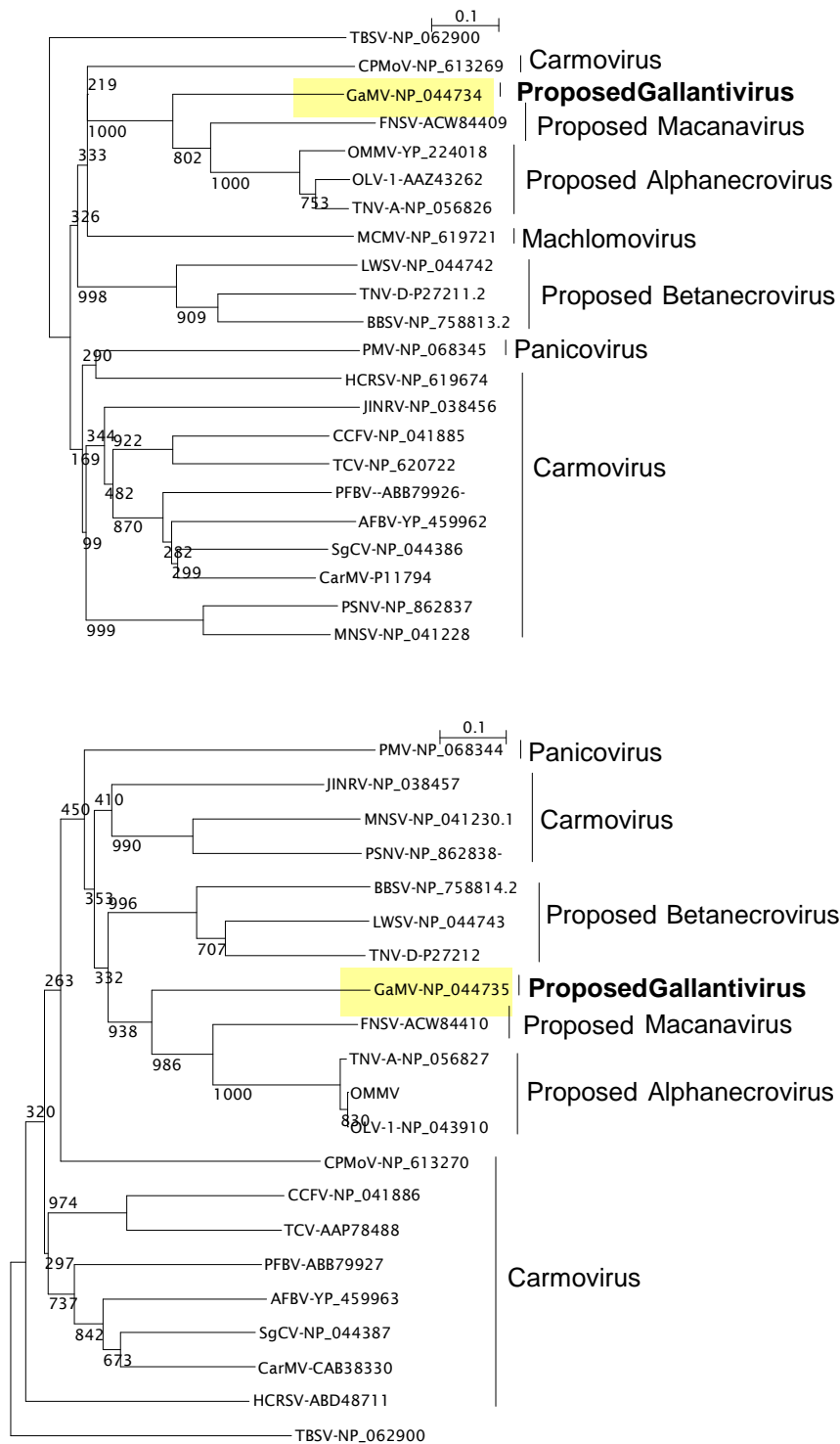
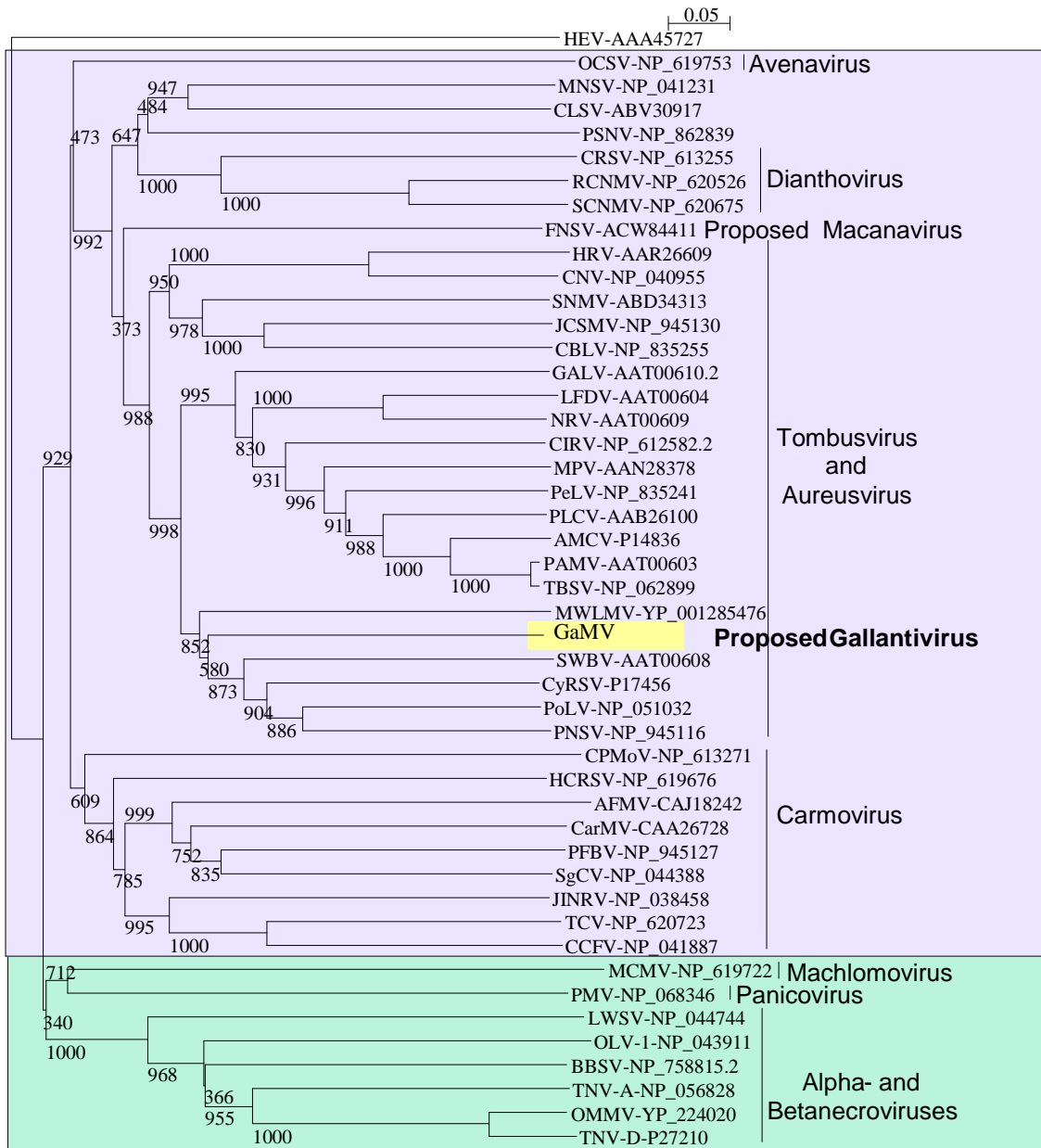


Figure 4. Phylogenetic (distance) tree of the MP1 (top) and MP2 (bottom) sequences of *Tombusviridae* members with the position of the GaMV MPs highlighted in yellow. Sequences were aligned using the ClustalX 2.1 algorithm and trees were generated by the Neighbor Joining method using 1000 bootstrap replicates.



- Virus coat proteins with protruding domains
- Virus coat proteins without protruding domains

Figure 5. Phylogenetic (distance) tree of CP sequences of *Tombusviridae* members with the position of the GaMV CP highlighted in yellow. Sequences were aligned using the ClustalX 2.1 algorithm and trees were generated by the Neighbor Joining method using 1000 bootstrap replicates. Hepatitis E virus (HEV) coat protein was used as an outgroup.

Table 1. Percent amino acid sequence identity between the polymerase of GaMV and those of the proposed alpha- and necrovirus genera and other *Tombusviridae* members

	% aa sequence identity in polymerase			
	GaMV	Alpha-necrovirus	Beta-necrovirus	Other Tombusviridae
GaMV	-	51	30-33	24-39

Table 1. Percent amino acid sequence identity between the MP1 of GaMV and those of the proposed alpha- and necrovirus genera and other *Tombusviridae* members

	% aa sequence identity in MP1					
	GaMV	Alpha-necro	Beta-necro	Carmo-	PMV	MCMV
GaMV	-	37-42	8-22	8-22	14	17

Table 1. Percent amino acid sequence identity between the MP2 of GaMV and those of the proposed alpha- and necrovirus genera and other *Tombusviridae* members

	% aa sequence identity in MP2				
	GaMV	Alpha-necro	Beta-necro	Carmo-	PMV-
GaMV	-	37	19-25	10-24	6

Table 1. Percent amino acid sequence identity between the CP of GaMV and those of the proposed alpha- and necrovirus genera and other *Tombusviridae* members

	% aa sequence identity in coat protein									
	GaMV	Alpha-necro	Beta-necro	PMV	MCMV	Tombus	Aureus	Carmo	OCSV	Diantho
GaMV	-	14-15	11-19	15	14	31-49	33-50	14-30	24	22-25

- Virus coat proteins with protruding domains
- Virus coat proteins without protruding domains

ABBREVIATIONS USED IN THE PROPOSAL

Tombusviruses

<i>Artichoke mottled crinkle virus</i>	AMCV
<i>Carnation Italian ringspot virus</i>	CIRV
<i>Cucumber Bulgarian latent virus</i>	CBLV
<i>Cucumber necrosis virus</i>	CNV
<i>Cymbidium ringspot virus</i>	CyRSV
<i>Pear latent virus - Italy</i>	PeLV*
<i>Grapevine Algerian latent virus</i>	GALV
<i>Havel river virus</i>	HRV
<i>Lato river virus</i>	LRV
<i>Moroccan pepper virus</i>	MPV
<i>Neckar river virus</i>	NRV
<i>Pelargonium leaf curl virus</i>	PLCV
<i>Pelargonium necrotic spot virus</i>	PNSV
<i>Petunia asteroid mosaic virus</i>	PAMV
<i>Sikte waterborne virus</i>	SWBV
<i>Tomato bushy stunt virus</i>	TBSV
<i>Limonium flower distortion virus</i>	LFDV

*PeLV is an isolate of *Eggplant mottle crinkle virus*

Dianthoviruses

<i>Carnation ringspot virus</i>	CRSV
<i>Red clover necrotic mosaic virus</i>	RCNMV
<i>Sweet clover necrotic mosaic virus</i>	SCNMV

Aureusviruses

<i>Cucumber leaf spot virus</i>	CLSV
<i>Johnsongrass chlorotic stripe mosaic virus</i>	JCSMV
<i>Maize white line mosaic virus</i>	MWLMV
<i>Pothos latent virus</i>	PoLV
<i>Sesame necrotic mosaic virus</i> (tentative species)	SNMV

Avenavirus

<i>Oat chlorotic stunt virus</i>	OCSV
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Carmoviruses

<i>Angelonia flower break virus</i>	AFBV
<i>Cardamine chlorotic fleck virus</i>	CCFV

<i>Carnation mottle virus</i>	CarMV
<i>Cowpea mottle virus</i>	CPMoV
<i>Galinsoga mosaic virus</i>	GaMV
<i>Hibiscus chlorotic ringspot virus</i>	HCRSV
<i>Japanese iris necrotic ring virus</i>	JINRV
<i>Melon necrotic spot virus</i>	MNSV
<i>Pea stem necrosis virus</i>	PSNV
<i>Pelargonium flower break virus</i>	PFBV
<i>Saguaro cactus virus</i>	SgCV
<i>Turnip crinkle virus</i>	TCV

Necroviruses

<i>Beet black scorch virus</i>	BBSV
<i>Leek white stripe virus</i>	LWSV
<i>Olive latent virus 1</i>	OLV-1
<i>Olive mild mosaic virus</i>	OMMV
<i>Tobacco necrosis virus A</i>	TNV-A
<i>Tobacco necrosis virus D</i>	TNV-D

Proposed Macanavirus

<i>Furcraea necrotic streak virus</i>	FNSV
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Panicovirus

<i>Panicum mosaic virus</i>	PMV
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Machlomovirus

<i>Maize chlorotic mottle virus</i>	MCMV
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Proposed Zeavirus

<i>Maize necrotic streak virus</i>	MNeSV
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